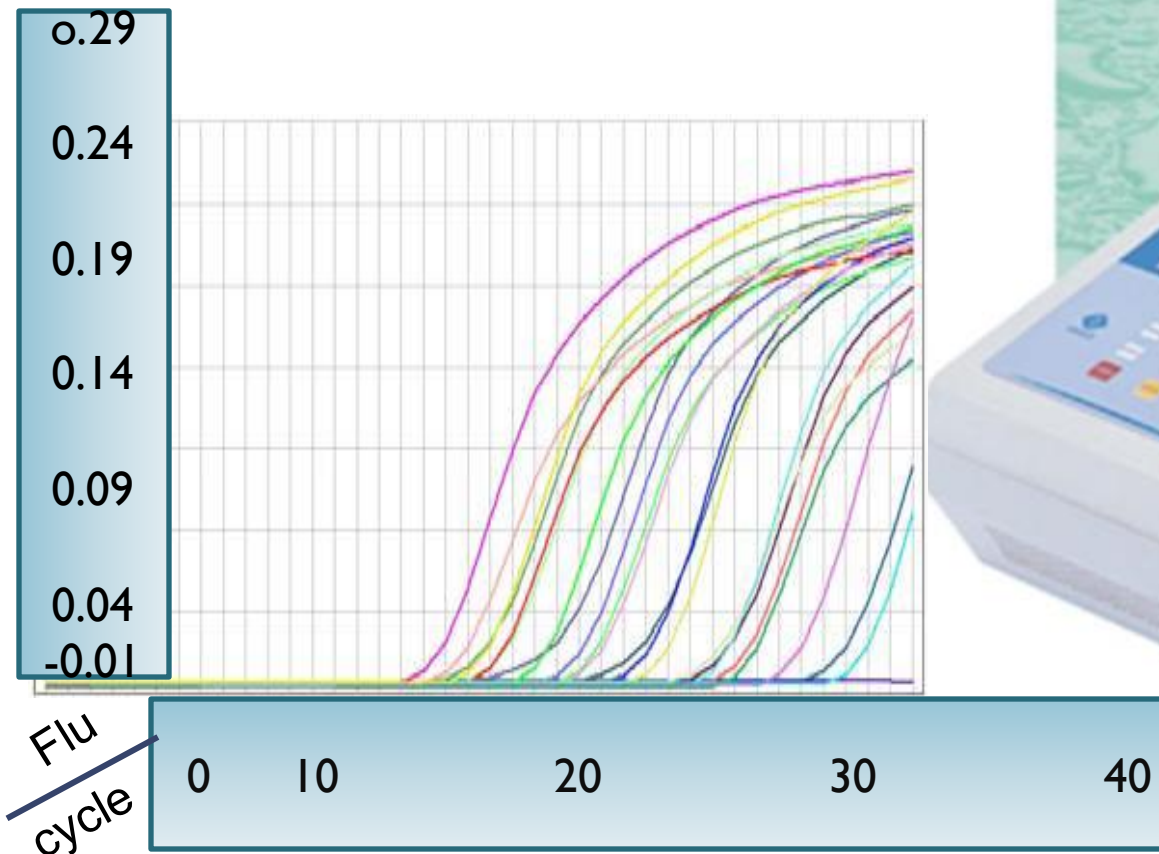


# REAL – TIME POLYMERASE CHAIN REACTION



There is three basic steps which are in common with all type of PCR

**Thermal denaturation :**

In this step DNAs are denatured mostly by temperature about 94° c & single stranded DNAs are made.

*( in some cases It's done by helicase )*

**Primer annealing :**

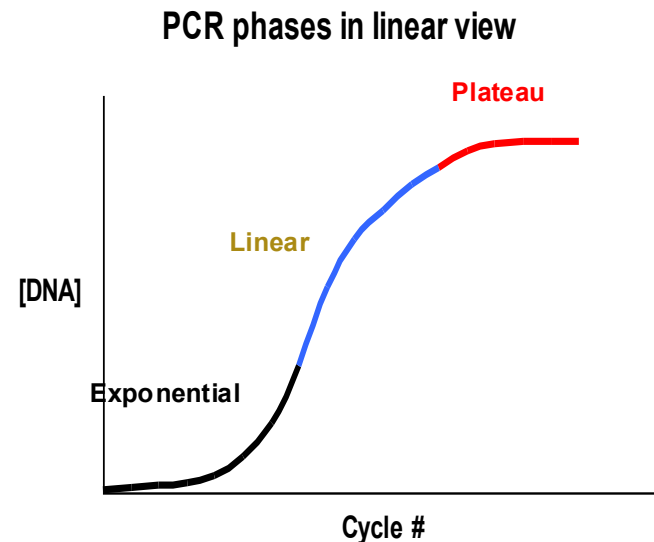
In this step Primers are attached to ssDNA by their complementary regions.

**Extension or polymerization :**

This is done by a temperature resistance polymerase named **Taq polymerase** which is extracted from **Thermus aquaticus**.

# PCR phases

- **Exponential**
  - If 100% efficiency – exact doubling of products. Specific and precise
- **Linear**
  - High variability. Reaction components are being consumed and PCR products are starting to degrade.
- **Plateau**
  - End-point analysis. The reaction has stopped and if left for long – degradation of PCR products.



# Advantages & disadvantages

- \* The most accurate & feasible technique to determine the amount & concentration of products.
- \* Rapid cycling (30 minutes to 2 hours).
- \* Specific & sensitive.
- \* Not much more expensive.

\* \* \* \* \*

- \* Pollution.
- \* Poor precision.
- \* Hard to get quantitative data.



# Real Time PCR

*Quantitative Real-Time PCR is an important technique for **quantifying** messenger RNA levels (gene expression) and DNA gene levels (copy number) in biological samples.*

Additional benefits of Real-Time quantitative PCR include sensitivity and a wide dynamic range. As few as 10 copies of an RNA/DNA target can be detected.

Considered to be the most **sensitive** method for the detection and quantification of gene expression

# Real Time -PCR

# What is Real Time-PCR ?

- Real-Time PCR is a specialized technique that allows a PCR reaction to be visualized “in real time” as the reaction progresses.
- This enables researchers to **quantify** the amount of DNA in the sample at the start of the reaction!
- It differs from standard PCR in a way that it can detect the amplified product as the reaction progresses with time but in standard PCR the amplified product is detected at the end of the reaction by agarose gel electrophoresis.

# Why Real Time? What's wrong with Agarose Gels?

- End point analysis
  - End point result is time consuming
  - End point is variable from sample to sample, while gels may not be able to resolve these variability's in yields.
- Low resolution
- Low sensitivity
- Size-based discrimination only

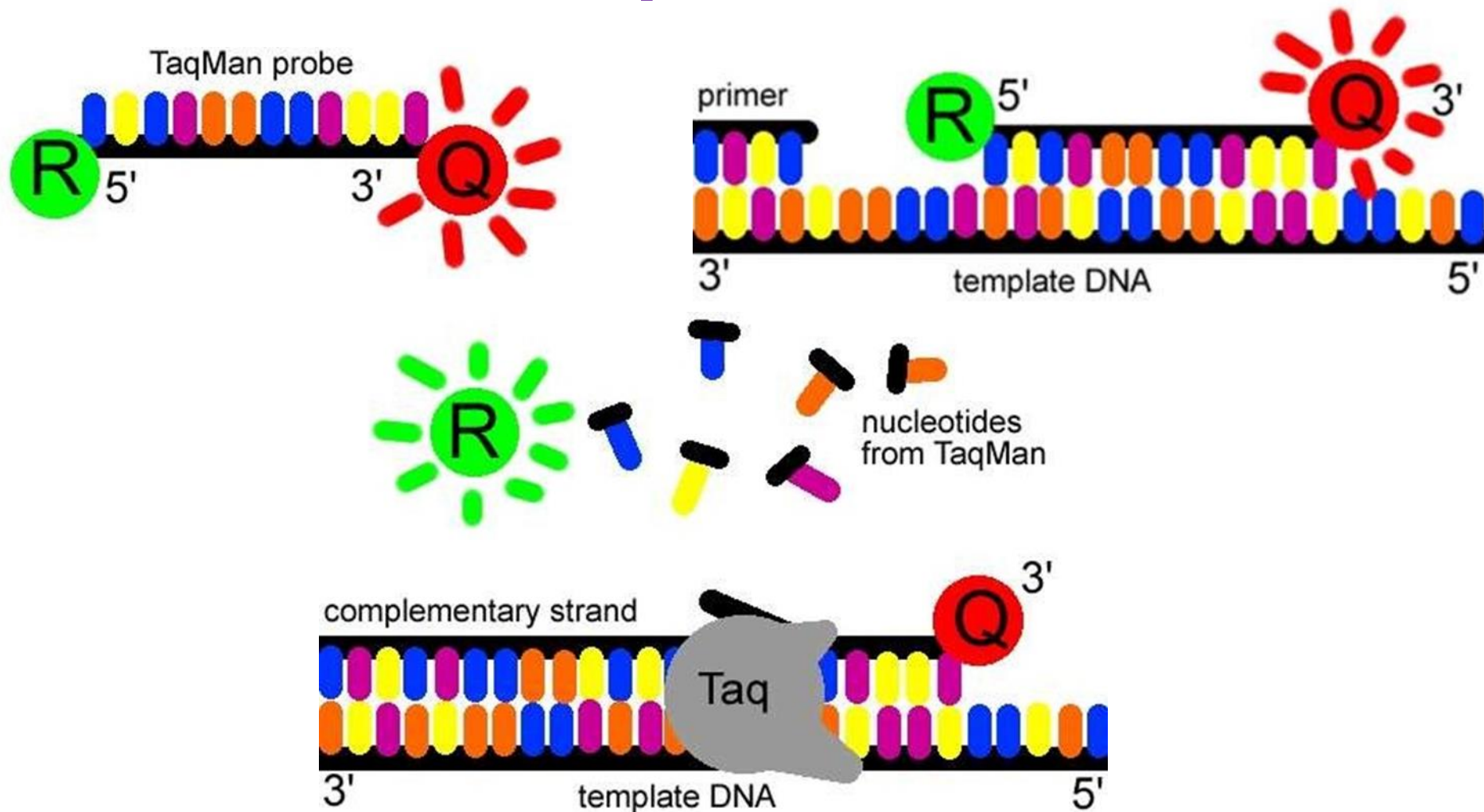
Real-time PCR enables the amount of starting material to be quantified



# Real Time Principle

- It is based on the detection and quantitation of a fluorescent reporter
- In stead of measuring the endpoint we focus on the first significant increase in the amount of PCR product.
- If there are only a few DNA molecules at the beginning of the PCR then relatively little product will be made, but if there are many starting molecules then the product yield will be higher.

# For Real Time PCR we need a specific probe with a fluorescent reporter





# Real-Time PCR

- Real-time PCR is a variant of PCR technology that allows the the detection of PCR products as they accumulate in "real-time" during the PCR amplification process.
- All real-time PCR systems rely upon the detection and quantitation of a fluorescent reporter  
The signal of which increases in direct proportion to the amount of PCR product in a reaction



## *A commonly misused acronym ...*

RT-PCR:

Reverse Transcription Polymerase Chain Reaction

- Or -

~~Real-Time Polymerase Chain Reaction?~~

RT-PCR = Reverse Transcription Polymerase Chain Reaction (may be applied to both conventional and Real-time PCR systems).

qPCR = Real-Time PCR when used as a quantitative tool.

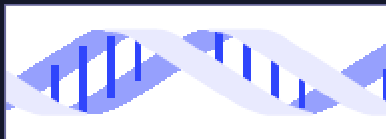
# An Overview of General PCR



**DNA  
Extraction**

**PCR**

**Visualization of  
PCR Products by  
Electrophoresis**

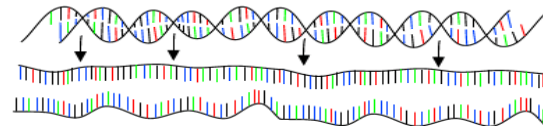


## PCR : Polymerase Chain Reaction

30 - 40 cycles

**Step 1 : d**

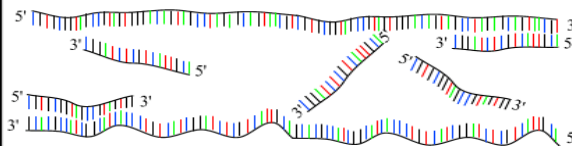
1 minut



**Step 2 : a**

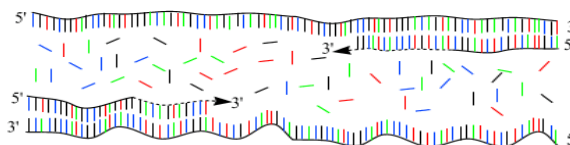
45 secon

forward  
primers



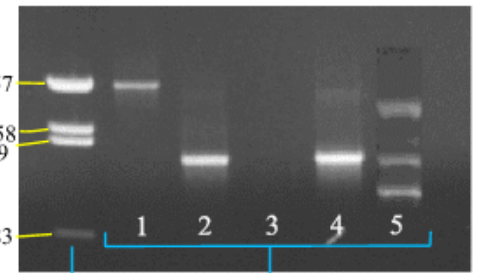
**Step 3 : extension**

2 minutes 72 °C  
only dNTP's



Verification of PCR product on  
agarose or searide gel

3kb  
1kb  
500  
300

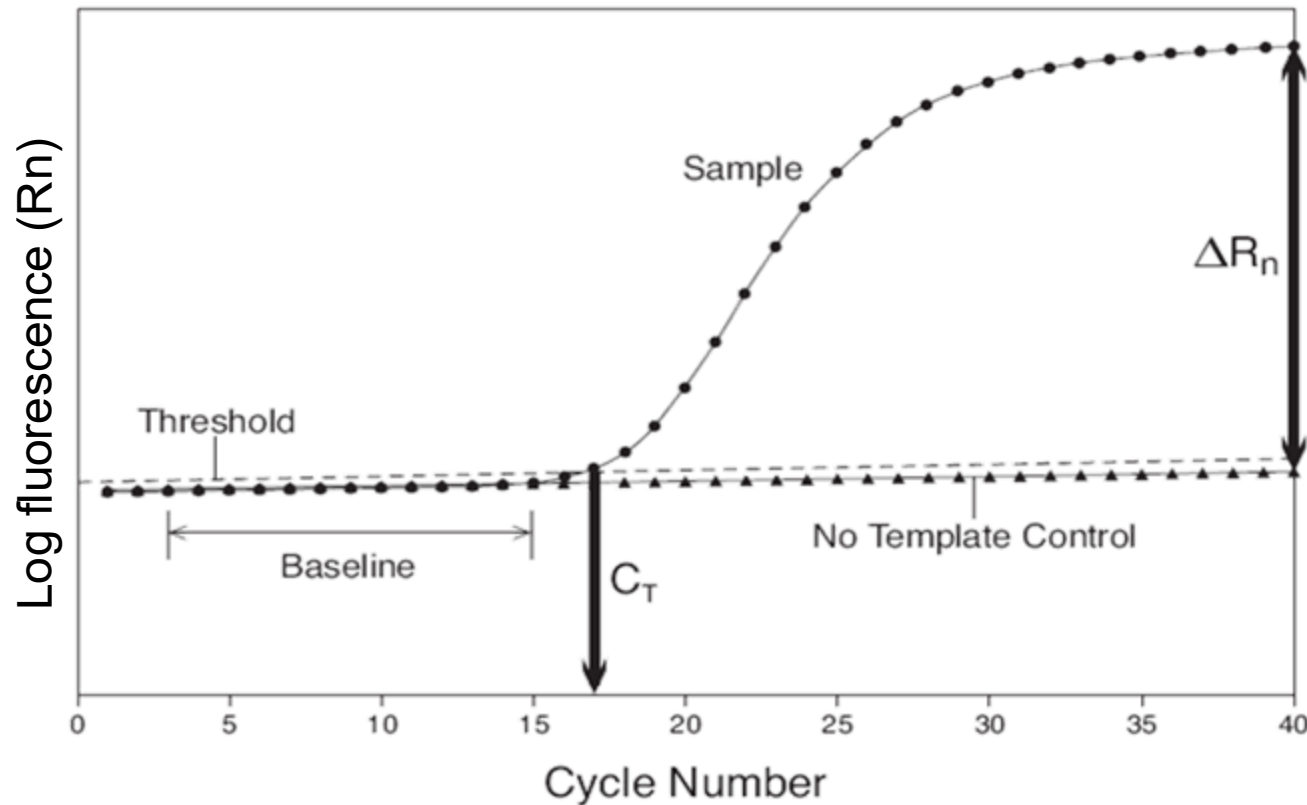


ladder

PCR fragments

(Andy Vierstraete 1999)

# The Basic of Real time PCR



**Baseline** - The baseline phase contains all the amplification that is below the level of detection of the real time instrument.

**Threshold** - where the threshold and the amplification plot intersect defines  $C_T$ . Can be set manually/automatically

$C_T$  - (cycle threshold) the cycle number where the fluorescence passes the threshold

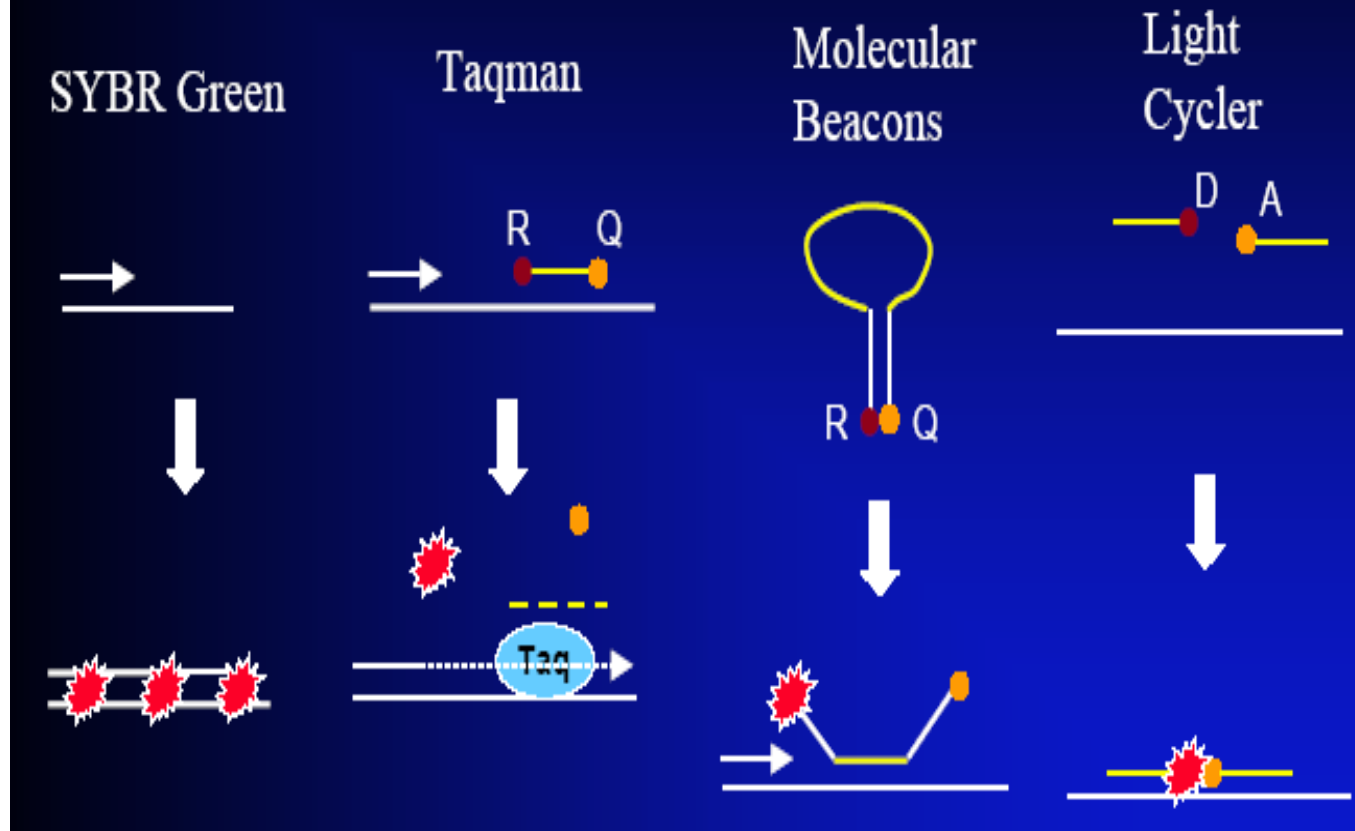
$\Delta R_n$  - ( $R_n$ -baseline)

**NTC** - no template control

$\Delta R_n$  is plotted against cycle numbers to produce the amplification curves and gives the  $C_T$  value.

# Detection in real time PCR

## Methods of fluorescence detection



# CYBR Green Chemistry



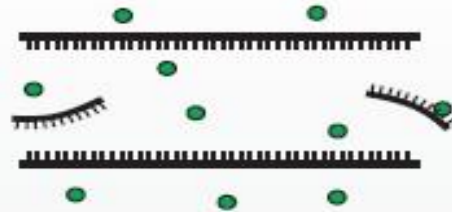
- ❑ CYBR Green is the most widely used double-strand DNA specific dye
- ❑ It binds to the minor groove of the DNA double helix
- ❑ In solution, the unbound dye exhibits very little fluorescence
- ❑ When CYBR Green dye binds to double stranded DNA, the fluorescent is substantially enhanced
- ❑ As more double stranded amplicons are produced SYBR green dye signal will increase



# SYBR<sup>®</sup> green

SYBR Green I fluoresces only when bound to dsDNA.

Denature



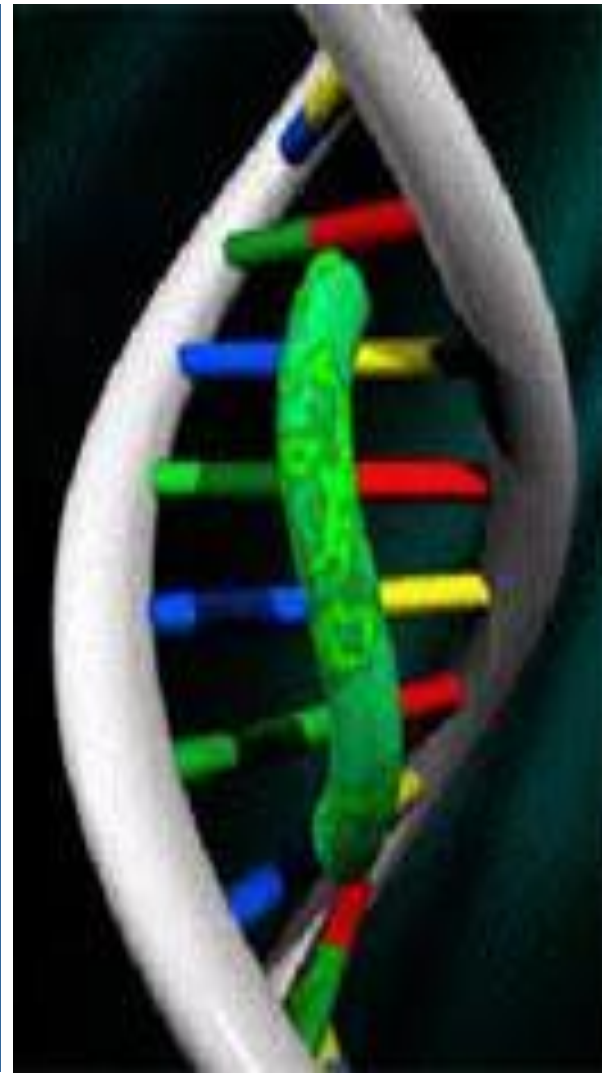
Anneal



Extend



- \* Pros: relatively cheap, doesn't require probe design
- \* Cons: nonspecificity can lead to false positives, not attuned for complex protocols



# Advantage and disadvantage of SYBR Green Method



## Advantage

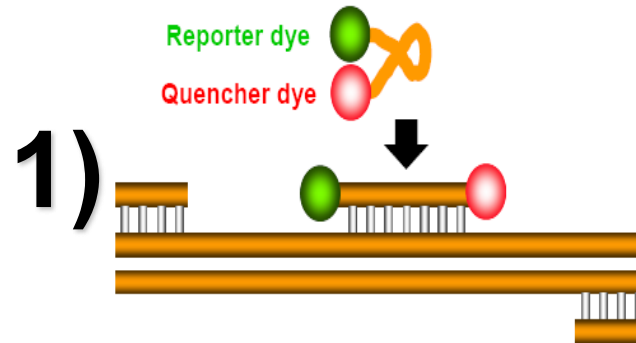
- ❑ Inexpensive
- ❑ No probe is required
- ❑ Easy to use

## Disadvantage

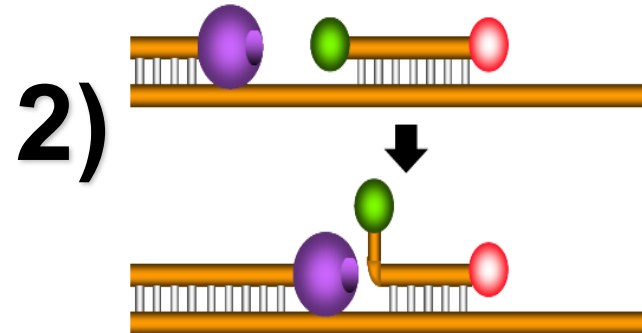
- ❑ SYBR Green will bind to any double stranded DNA (e.g. primer dimers, non-specific reaction products)
- ❑ Overestimation of target concentration
- ❑ Non-specific background in very late cycles

# Taqman

1) Denaturation and hybridization of probe.

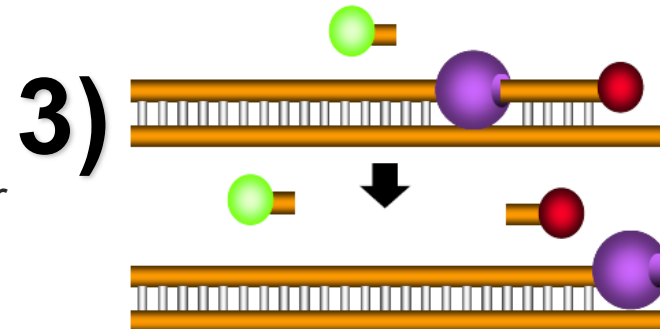


2) Extension of primer and strand displacement of probe.



3) Cleavage of probe and fluorescence from the reporter dye.

Fluorescence from reporter dye is directly proportional to the number of amplicons generated





# TaqMan Chemistry

TaqMan chemistry requires two more additional components with traditional PCR

## What's in general PCR ?

- ❑ Template DNA
- ❑ Reaction buffer
- ❑ Nucleotides (dNTPs)
- ❑ Primers (Forward & Reverse)
- ❑ *Taq* DNA polymerase

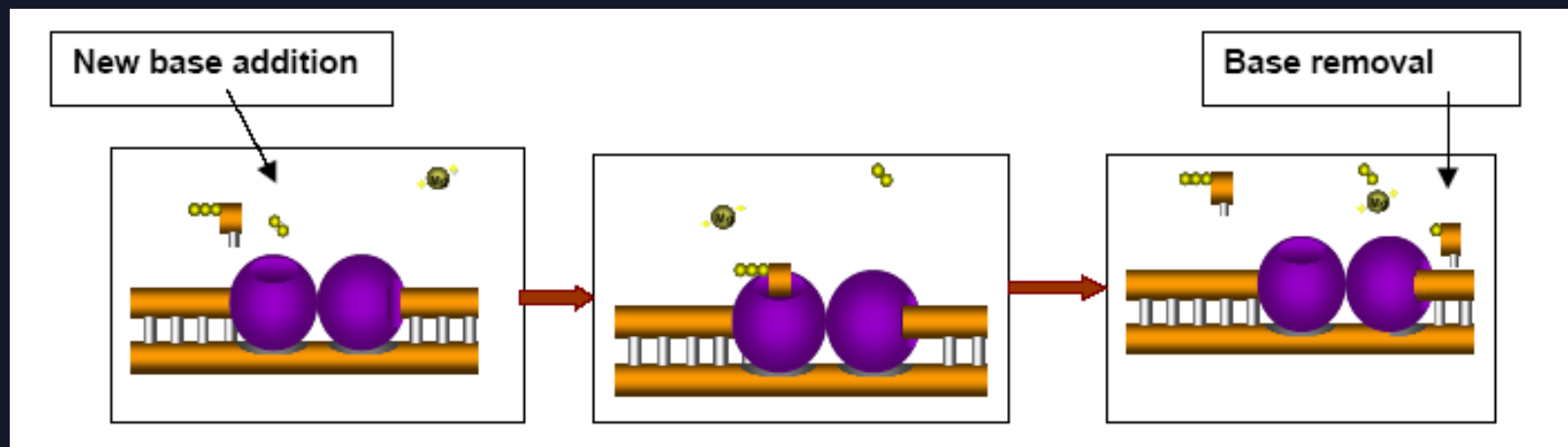
## What's new in TaqMan ?

- ❑ Template DNA
- ❑ Reaction buffer
- ❑ Nucleotides (dNTPs)
- ❑ Primers (Forward & Reverse)
- ❑ *AmpliTaq Gold* DNA Polymerase
- ❑ TaqMan Probe
- ❑

# AmpliTaq Gold DNA Polymerase



- ❑ In addition to its polymerase activity, it has also 5' exo-nuclease activity
- ❑ The 5' exo-nuclease activity acts upon the surface of the template to remove obstacles downstream of the growing amplicon that may interfere with its generation



# TaqMan Probe



- ❑ TaqMan probe is a short DNA sequence with a high energy dye called **reporter dye** at the 5' end and a low energy dye called **quencher** at the 3' end



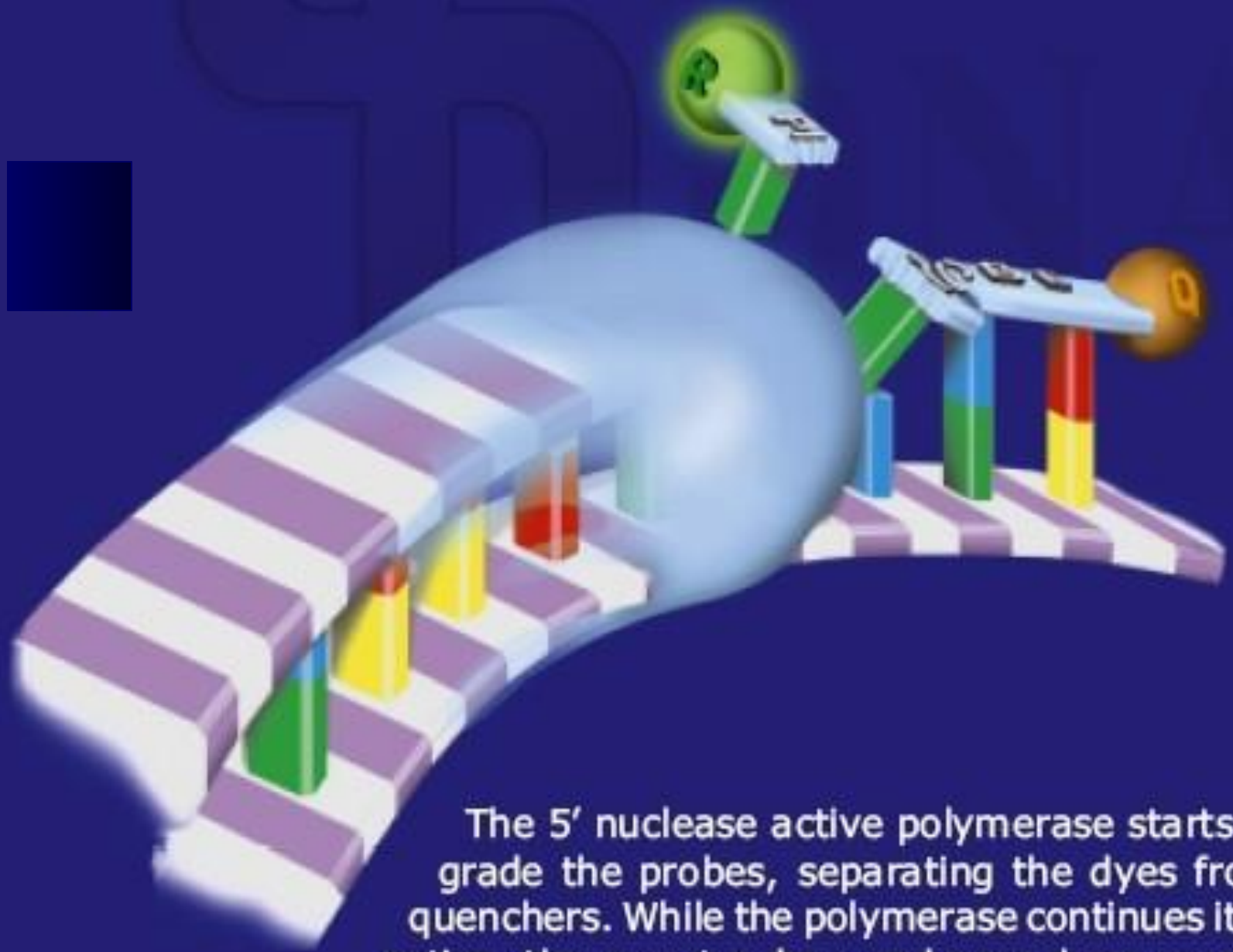
- ❑ When this probe is intact and excited by a light source, the reported dye emission is suppressed by the quencher dye as a result of close proximity of the dyes. This is known as **FRET**

# Fluorescence Resonance Energy Transfer



**When a high energy dye is in close proximity of a low energy dye, there will be a transfer of energy from high to low**





The 5' nuclease active polymerase starts to degrade the probes, separating the dyes from the quenchers. While the polymerase continues its elongation, the reporter dyes and quenchers are drifting apart, leaving the quenchers unable to absorb fluorescence emitted from the dyes. The result is a detectable fluorescence signal from the sample.



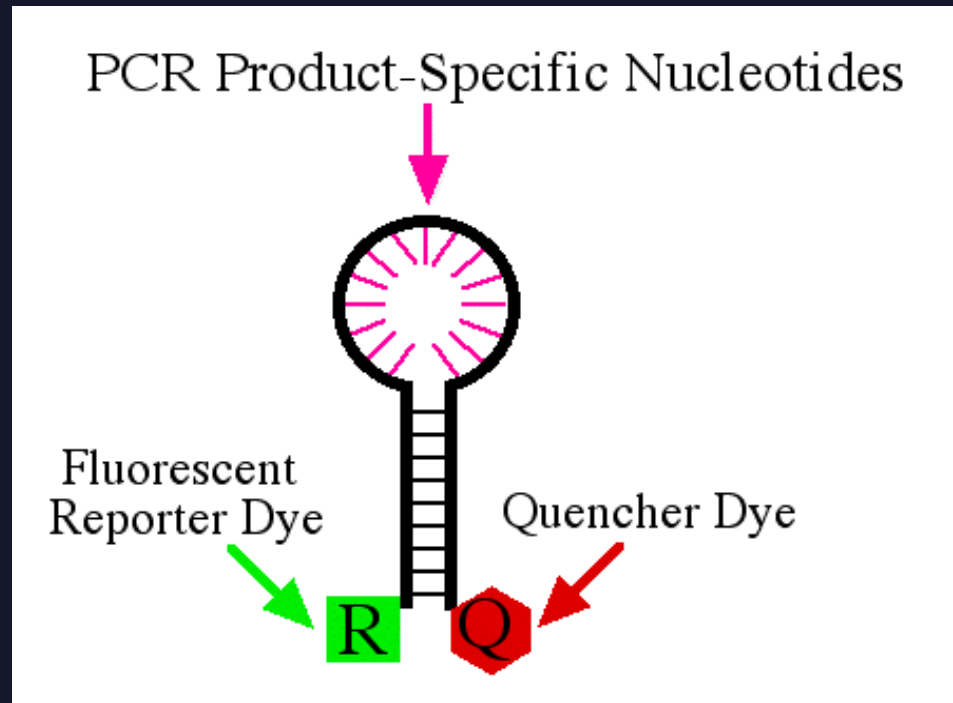


# Molecular Beacon

- Molecular beacons are short segments of single stranded DNA that forms a **hairpin** in its free form
- The loop portion of the molecular beacon is composed of bases that are complementary to one strand of the PCR product the investigator wants to detect and quantify
- Attached to opposite end of the beacon are a fluorescent reported dye and a quencher dye

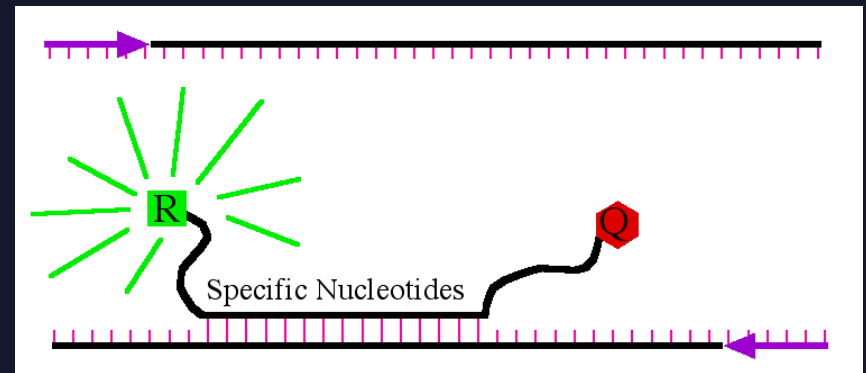
# Molecular Beacon

When the molecular beacon is in the hairpin conformation, any fluorescence emitted by the reporter is absorbed by the quencher dye and no fluorescence is detected.



# Molecular Beacon

- As the PCR continues, the newly synthesized PCR products are denatured by high temperatures
- At the same time the molecular beacon also is denatured so the hairpin structure is disrupted.
- As the temperatures cool for the next round of primer annealing, the molecular beacon is capable of forming base pairs with the appropriate strand of the PCR product





# Molecular Beacon

- ❑ Molecular beacons that bind to the PCR product remove the ability for the quencher to block fluorescence from the reporter dye
- ❑ Molecular beacons that do not bind to the PCR product reform the hairpin structure and thus unable to fluoresce

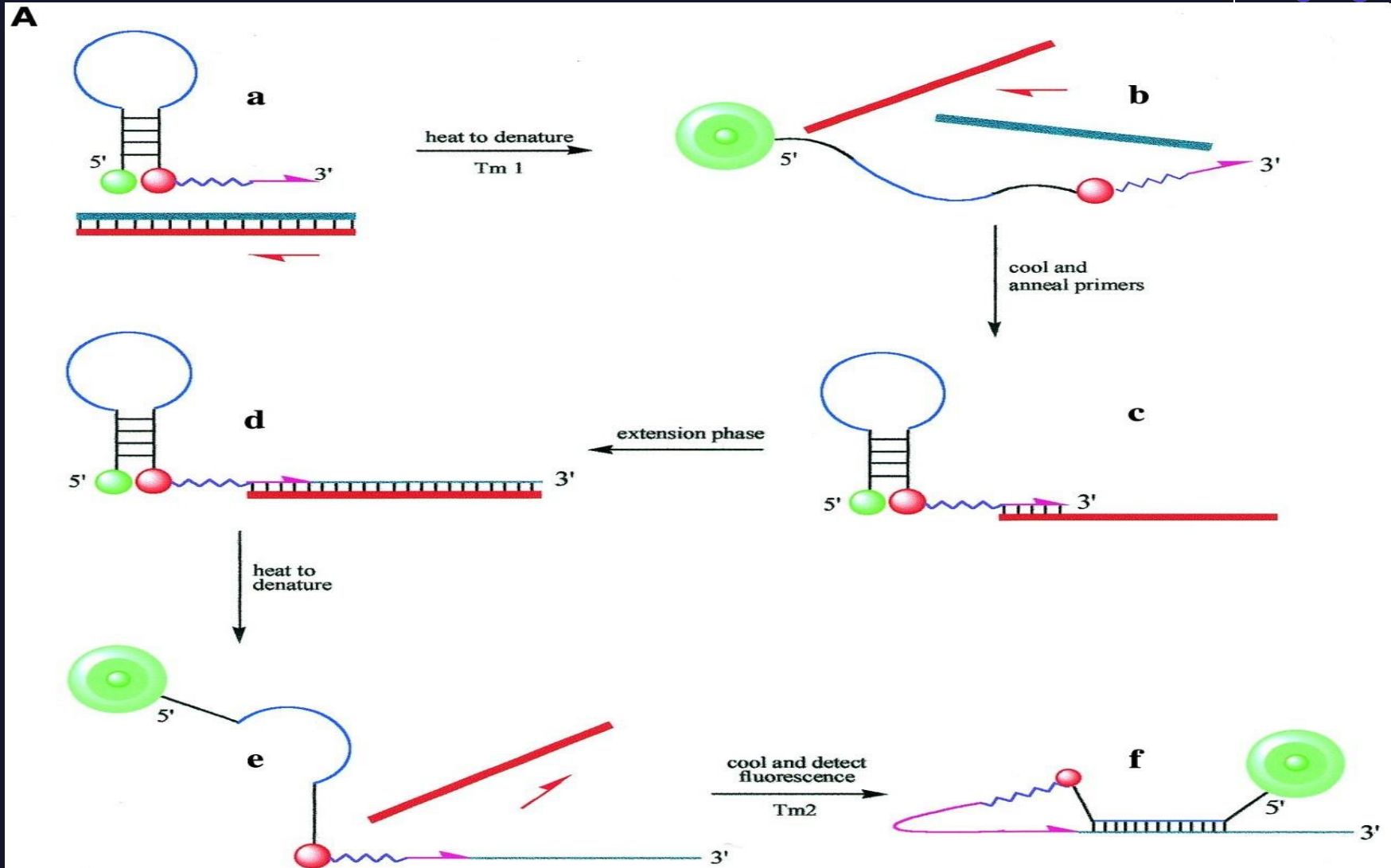
❑ Therefore, as PCR product accumulates, there is a linear increase in fluorescence.

# Scorpion probe



- ❑ Scorpion probe is a bifunctional molecule in which a primer is covalently linked to the probe
- ❑ That is why they are sometimes known as “**Scorpion primer & probe**”
- ❑ The probe has a self complementary stem sequence with a fluorophore at one end and a quencher at the other end
- ❑ In the initial PCR cycle the primer hybridizes to the target and extension occurs due to the action of polymerase
- ❑ After denaturation and cooling, the specific probe sequence is able to bind to its complement within the extended amplicon thus opening up the hairpin loop
- ❑ The fluorescent dye and quencher are separated, FRET does not occur, and the fluorescent dye emits light upon irradiation

# Scorpion primer & probe



# **Applications of Real Time PCR**

- **Quantitation of gene expression**
- **Drug therapy efficacy / drug monitoring**
- **Viral quantitation**
- **Pathogen detection**

