POLYMERASE CHAIN REACTION

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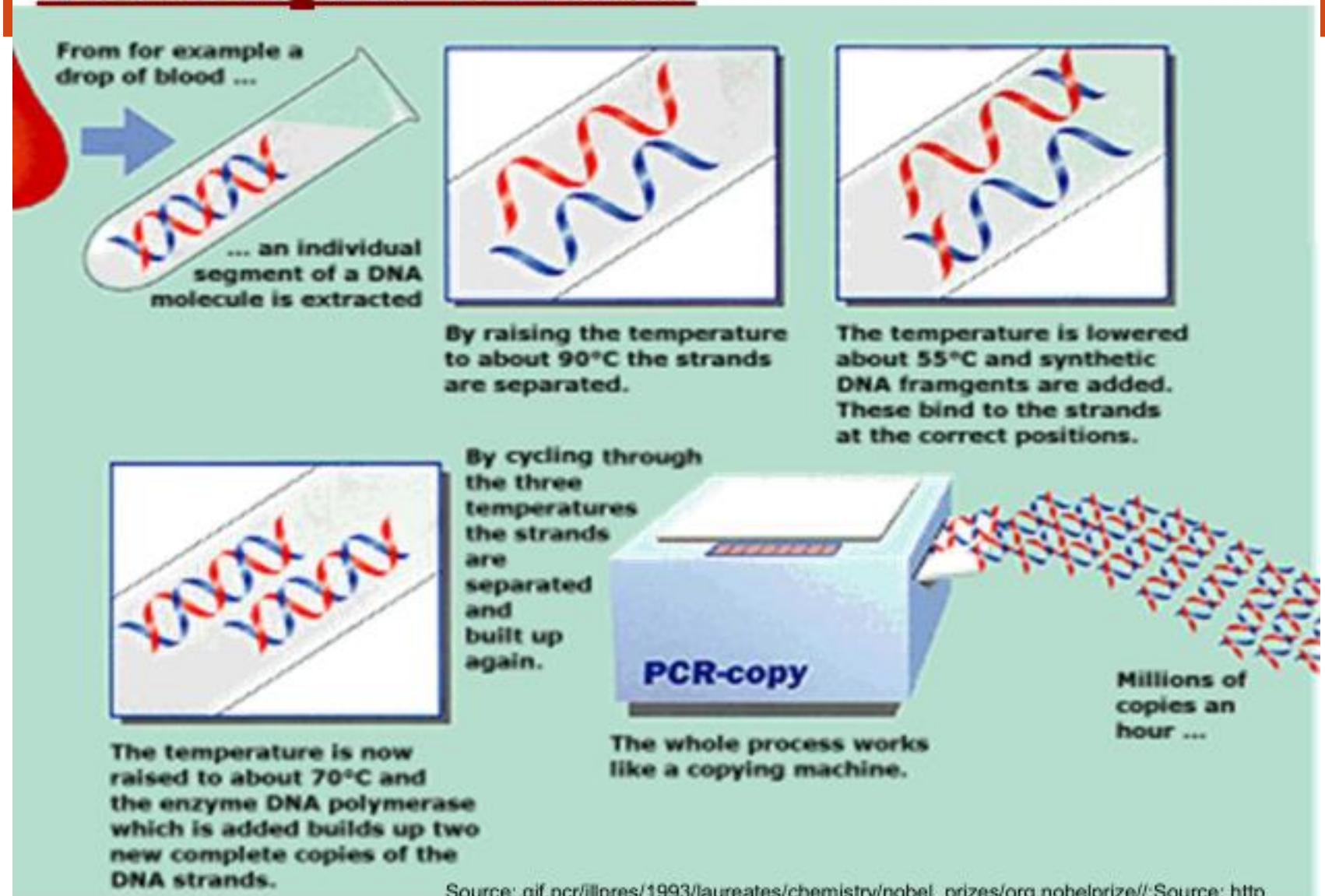
Nobel prize 1993



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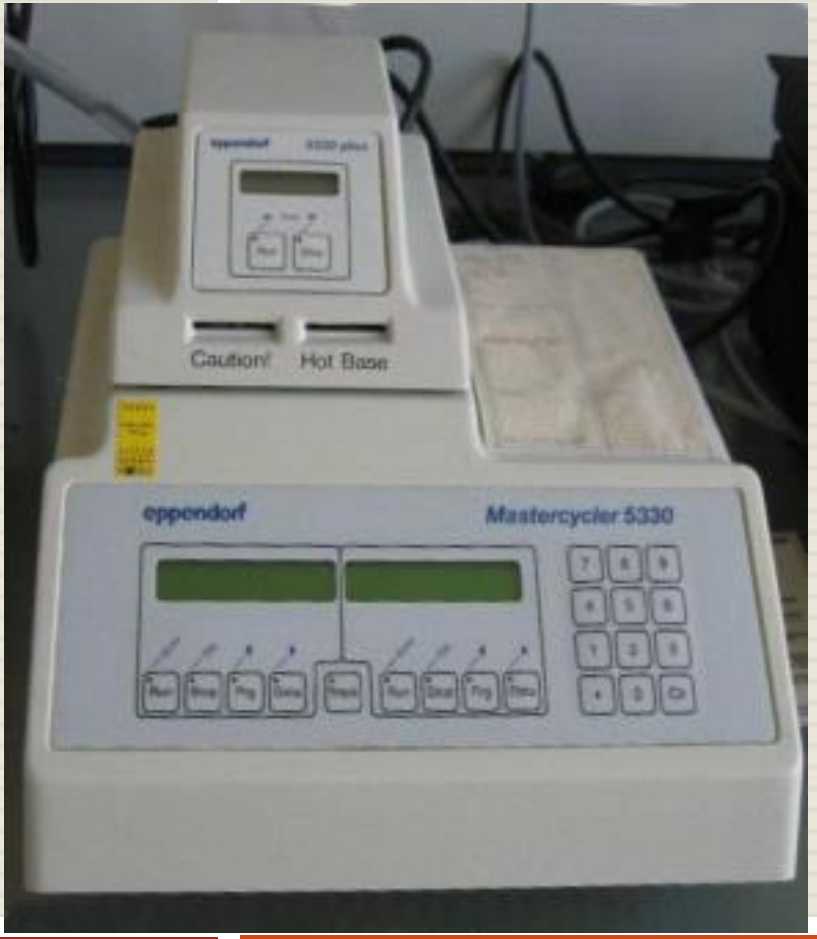
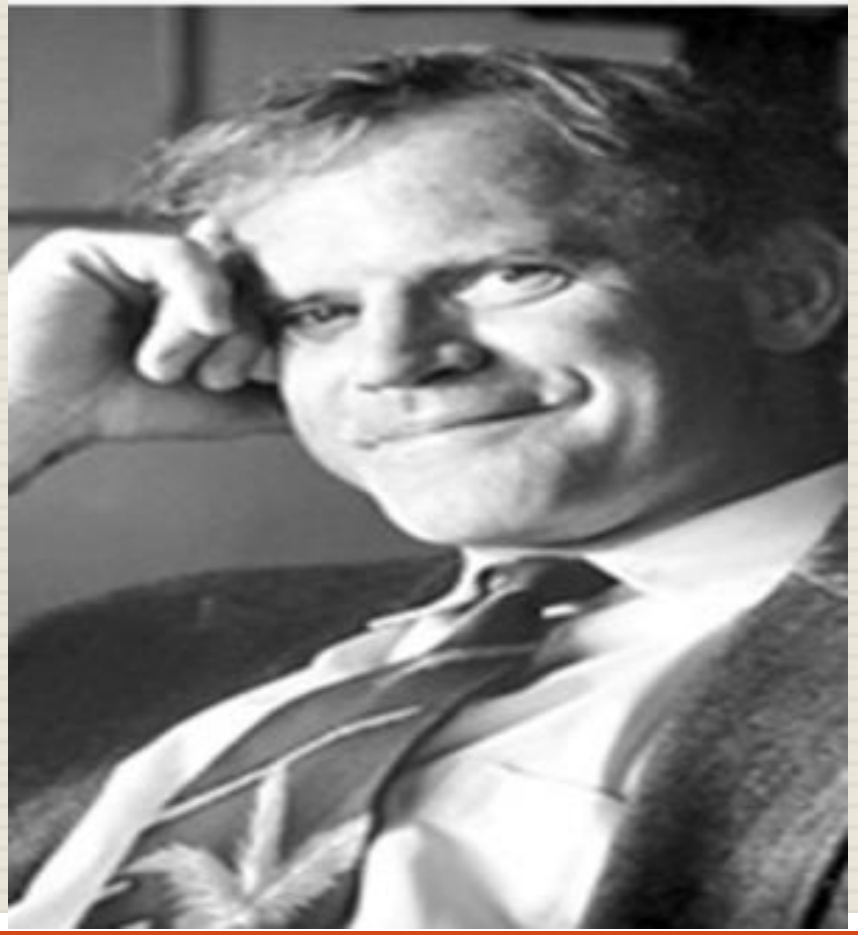


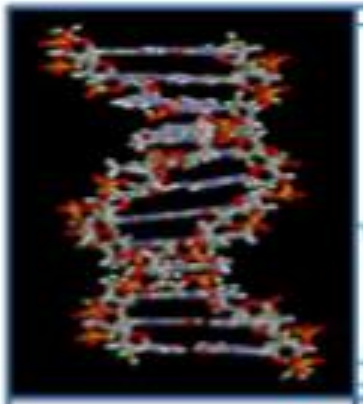
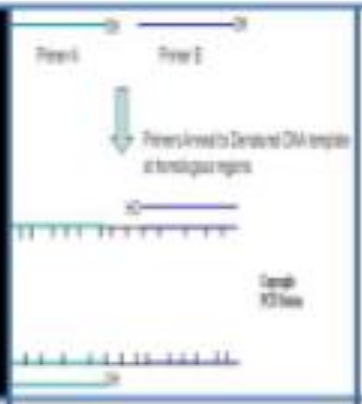


- Coping Machine for DNA Molecule
- Invented by Kary Mullis and his colleagues in the 1983

Polymerase Chain Reaction

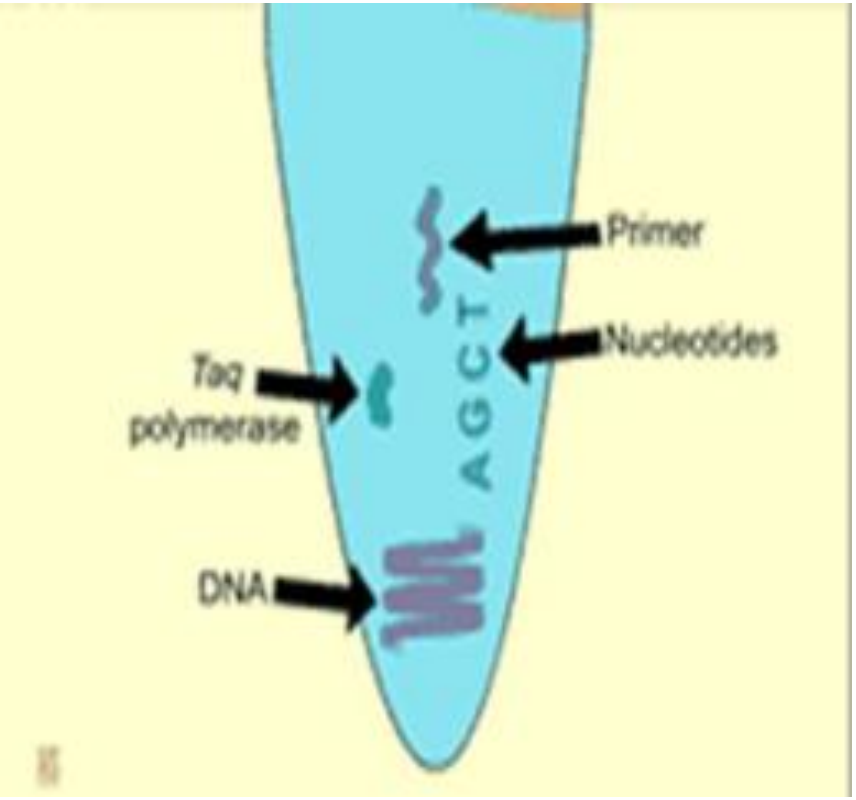
- **PCR:** Technique for in vitro (test tube) amplification of specific DNA sequences via the temperature mediated. DNA polymerase enzyme by simultaneous primer extension of complementary strands of DNA.
- **PCR:** This system for DNA replication that allows a "target" DNA sequence to be selectively amplified, several million-fold in just a few hours.

PCR reaction components

				
Template	Two primers (forward and reverse)	Four (dNTPs) A, G, C, T	Buffer system containing magnesium Mg^{2+}	DNA polymerase

PCR reaction components

- DNA template
- Two primers
- Four normal deoxynucleosides triphosphates
- Buffer system
- DNA polymerase I



DNA Template

Integrity

- High molecular weight

Purity

- Pure

Amount

- Human genomic DNA should be up to 500ng
- Bacterial DNA 1-10ng
- Plasmid DNA 0.1-1ng

Primers

- Typical primers are 18-28 bases in length,
- Having 40- 60% GC composition,
- Have a balanced distribution of G/C and A/T rich domains,
- The calculated Tm for a given primer pair should be balanced (difference no more than 5 °C),
- Tm=Temperature at which 2 strands of the duplex dissociate
- Primer concentration between 0.1 and 0.6 µM are generally optimal,
- Have a cytosine and guanine at the 3'-end because they form three hydrogen bonds with the matrix molecules, making a more stable hybridization

Calculation of Melting Temperature

Tm= 2 C° X (number of A and T bases)+4 C°X (number of G and C bases).

Optimal annealing temperature are 5-10 C ° lower than Tm values of the primers .

Four Normal Deoxynucleosides Triphosphate

- Final concentration of dNTPs should be 50-500 µM (each dNTP). Usually included at conc. of 200 µM for each nucleotide.
- Always use balanced solution of all four dNTPs to minimize polymerase error rate.

Buffer System Containing Magnesium

The standard PCR buffer contains:

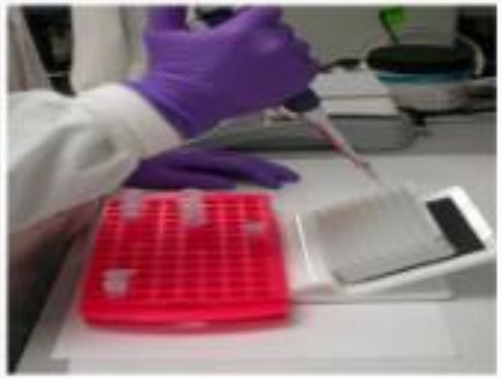
- **Tris-HCl 10mM (10-50mM)** for dissolution of nucleic acids
  pH 8.3 (pH 8.3-8.8 at 20C°)
- **KCl 50mM** promotes specificity of hybridization
- **MgCL2 1.5mM (0.5-10mM)** for stabilizing of complex between primers and matrix and for increasing of exit the special product of PCR
- **Gelatin or Bovine Serum Albumin 100 µg/ml**

frequent unfreezing-freezing at the temperature -20C

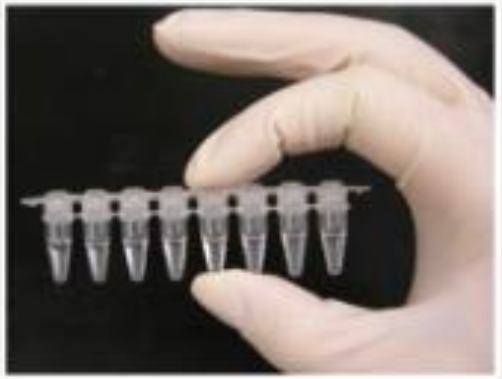
DNA Polymerase

- The most widely characterized polymerase is that from Thermus aquaticus (Taq), Thermophilic bacterium lives in hot springs and capable of growing at 70 -75 C°,
- Consist of a single polypeptide chain has a molecular weight of 95 Kd, and has an optimum polymerization temperature of 70 – 80 C° (72 C°).
- 0.5 – 2 units/50µl reaction. Too little will limit the amount of products, while too much can produce unwanted non specific products.

PCR



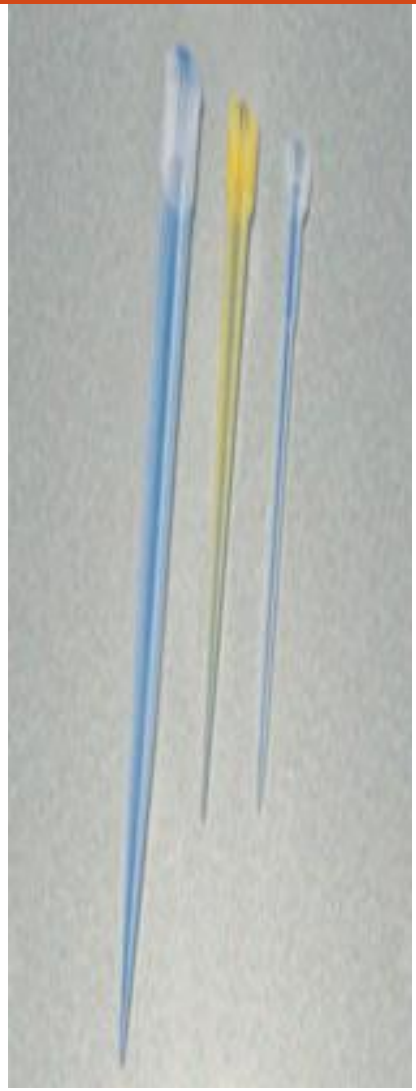
All one has to do is to mix the contents in the test tube and



Seal the tube and then place it in a thermal cycler.

Thermal cycler

AVOIDING CONTAMINATION



1-Sample Handling

- Use sterile techniques and always wear fresh gloves,
- Always use new or sterilized glassware, plasticware and pipettes to prepare the PCR reagents and template DNA,
- Autoclave and sterilize all reagents and solution,
- Have your own set of PCR reagent and Solution (store in small aliquots),
- Positive and negative control should be included.

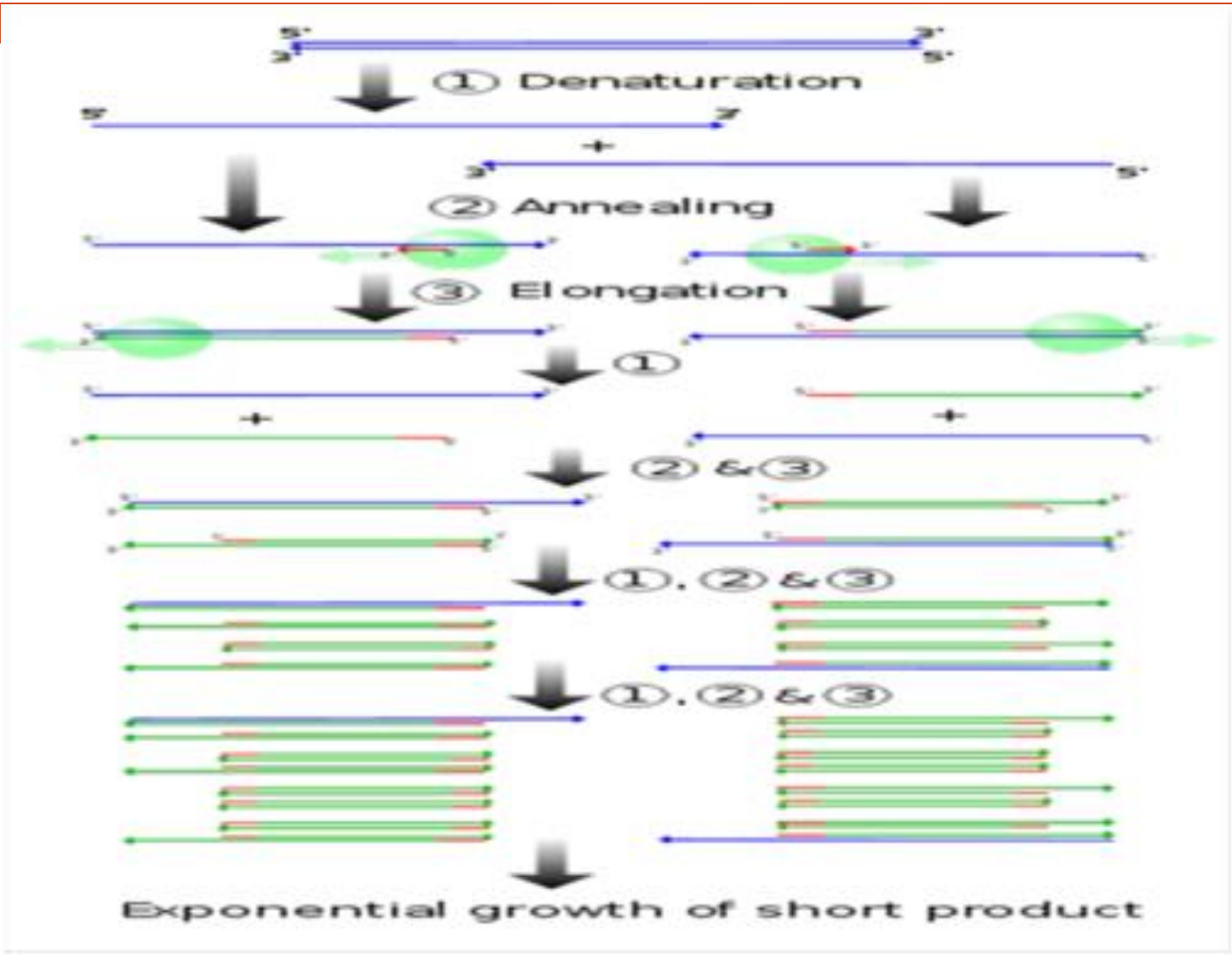
2-Laboratory Facilities

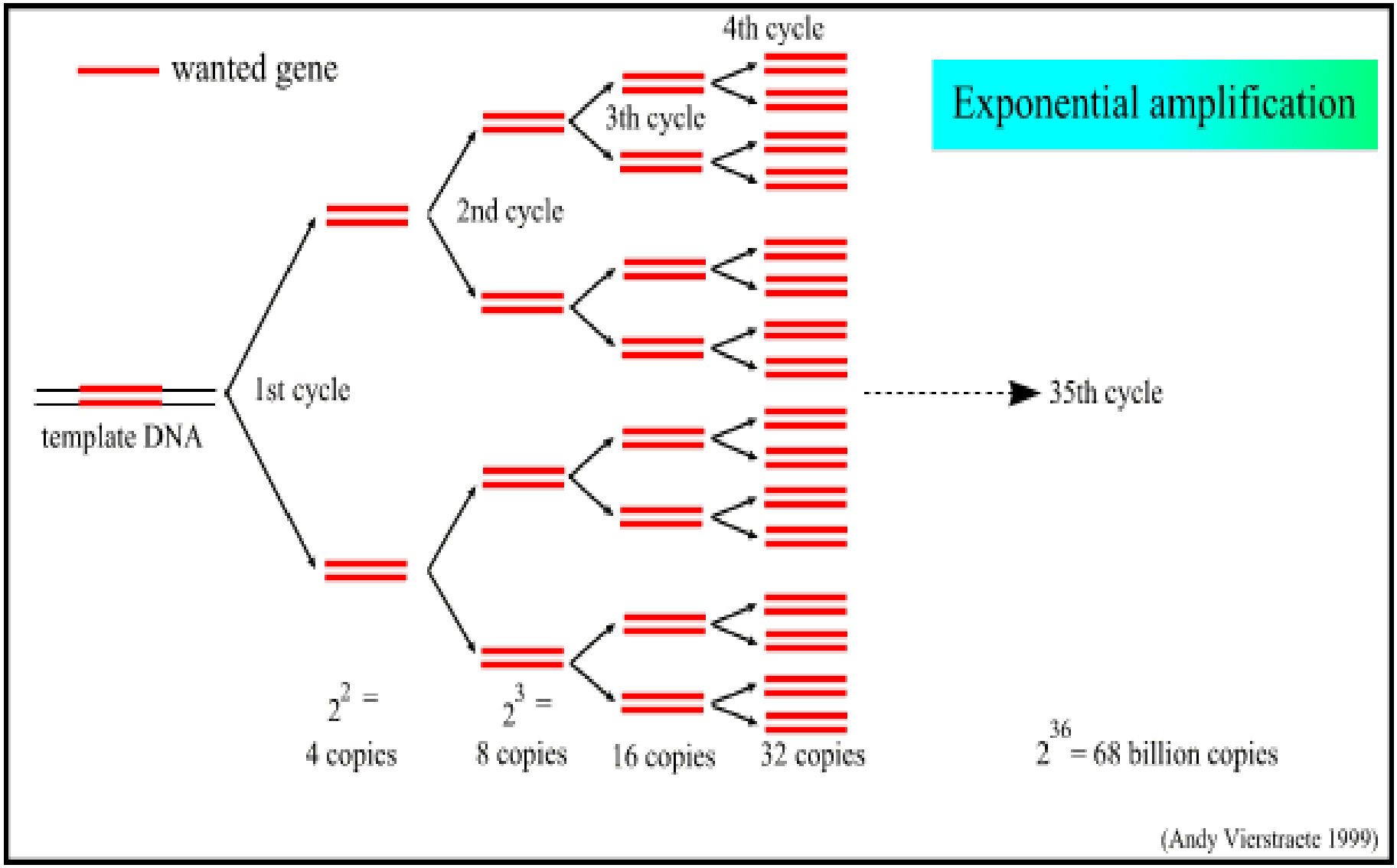
- Set up physically separated working places for:
  - Template preparation
  - Setting up PCR reactions
  - Post PCR analysis
- Use PCR only pipettes, micro-centrifuges and disposable gloves
- Use aerosol resistant pipette tips
- PCR reaction under a fume hood equipped with UV LIGHT.

3-Working with RNA

- Do not touch a surface after putting the gloves to avoid reintroduction of RNAse to decontaminated material.
- Designate a special area for RNA work only.
- Treat surface or benches and glassware with commercially available RNAse inactivating agents.

Polymerase Chain Reaction




wanted gene
Exponential amplification
4th cycle
3th cycle
2nd cycle
1st cycle
template DNA
35th cycle
2^2 =
4 copies
2^3 =
8 copies
16 copies
32 copies
2^{36} = 68 billion copies
(Andy Vierstracte 1999)

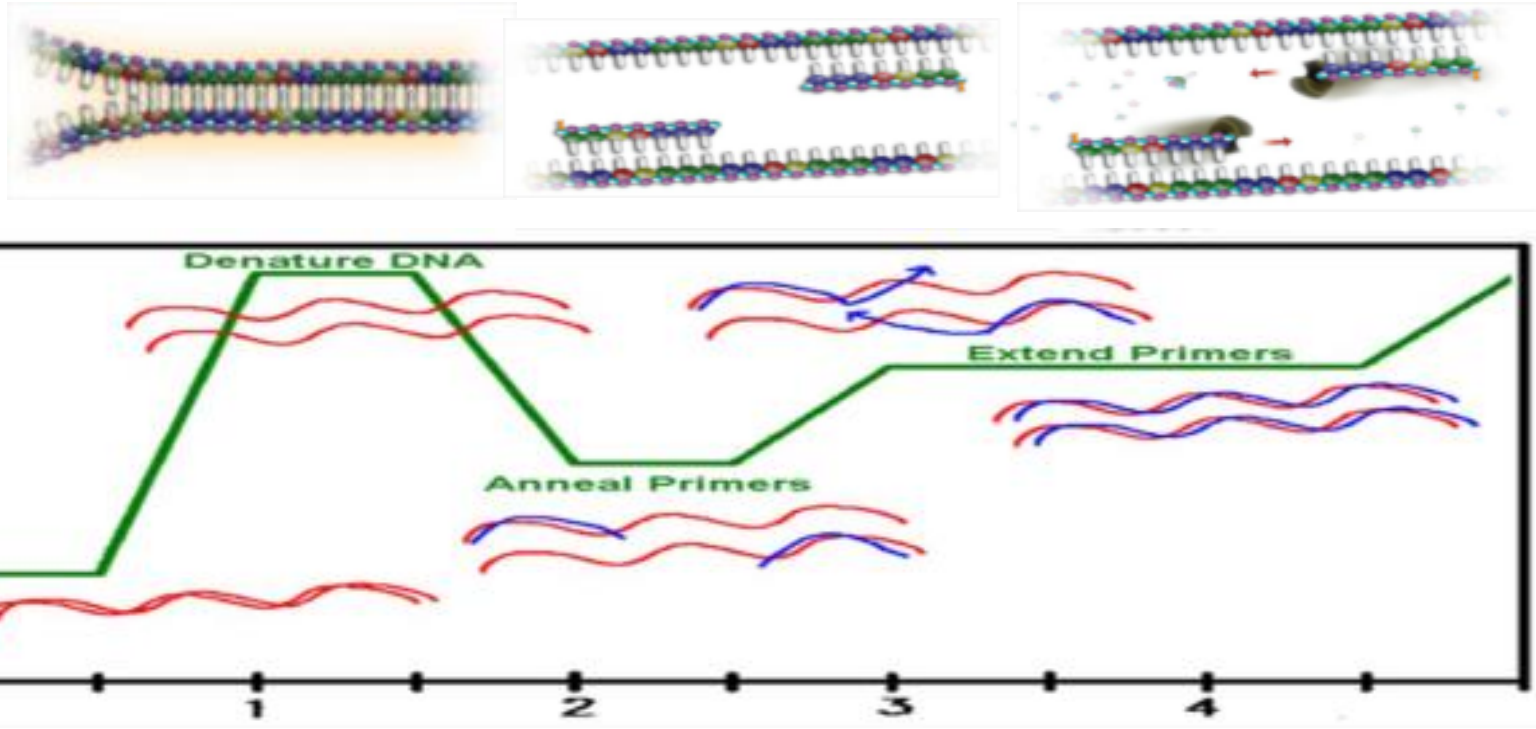
Thermal Cycling Profile for Standard PCR

- **Initial Denaturation:**
- Initial heating of the PCR mixture at 94- 95C within 2 min. is enough to completely denature complex genomic DNA.
- **Each cycle includes three successive steps:** Denaturation, annealing and extension.
- **Post extension and holding:**
- Cycling should conclude with a final extension at 72 C° for 5 -15 minute to promote completion of partial extension products and then holding at 4 C°.

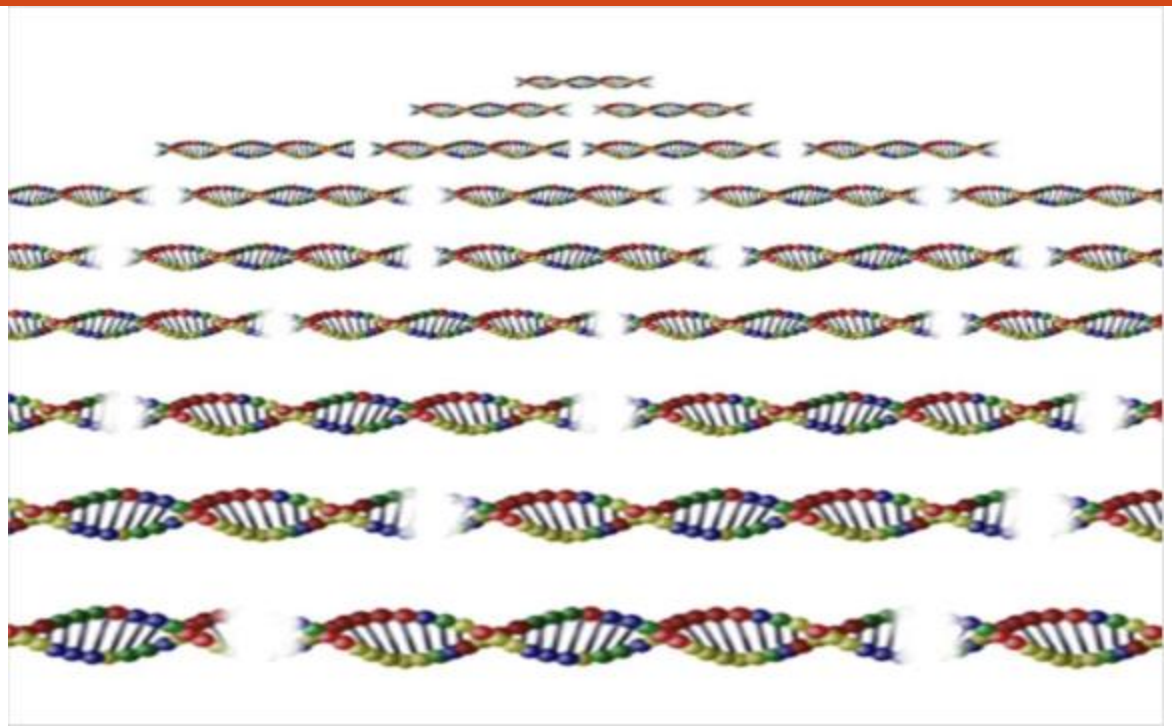
Each cycle includes three successive steps:

Denaturation	94C° 15sec-one minute	The DNA is denatured into single strands.
Annealing	34-72C° 30sec-two minute	The primers hybridize or "anneal" to their complementary sequences on either side of the target sequence.
Extension	72C° 1.5-3 minute	The polymerase binds and extends a complementary DNA strand from each primer

PCR


Denature DNA
Anneal Primers
Extend Primers
Temperature (°C)
100
90
80
70
60
50
40
30
20
10
0
Time
0
1
2
3
4

Exponential Amplification



As amplification proceeds, the DNA sequence between primers doubles after each cycle.

(The amplification of the target sequence proceeding in an exponential fashion (1 2 4→8→16→……) up to million of times the starting amount until enough is present to be seen by gel electrophoresis.

Number of Cycles

- The number of cycles required for optimum amplification varies depending on the amount of the starting material.

- Most PCR should, therefore, include only 25 – 35 cycles. As cycle increases, nonspecific products can accumulate.

- After 20- 40 cycles of heating and cooling build up over a million copies of original DNA molecules.

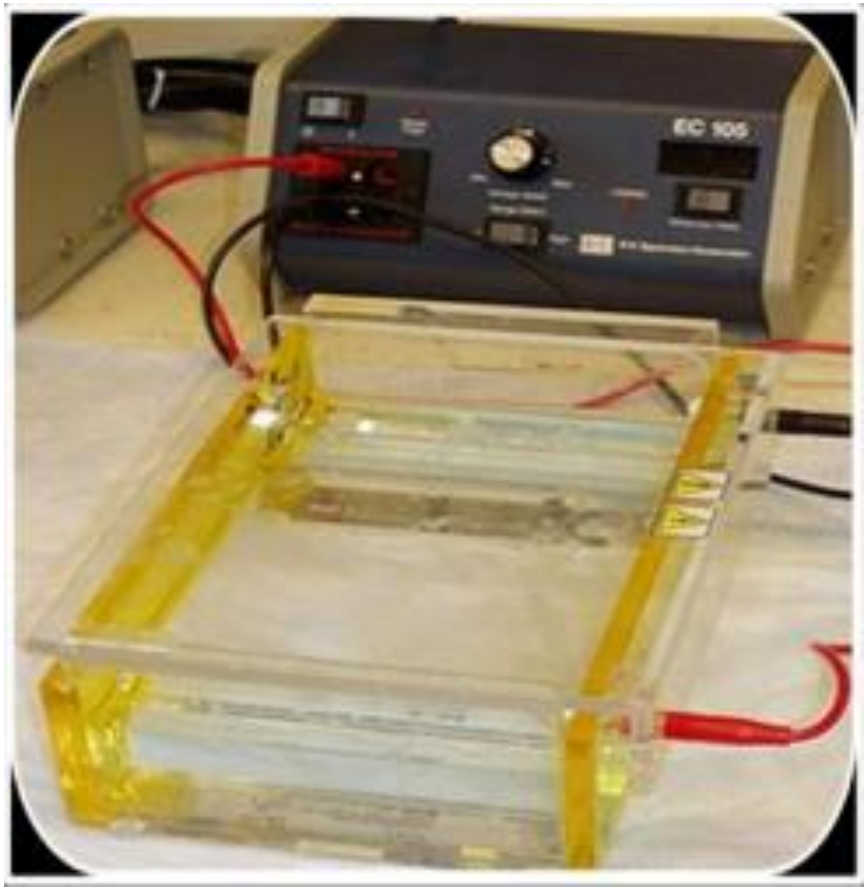
GEL ELECTROPHORESIS

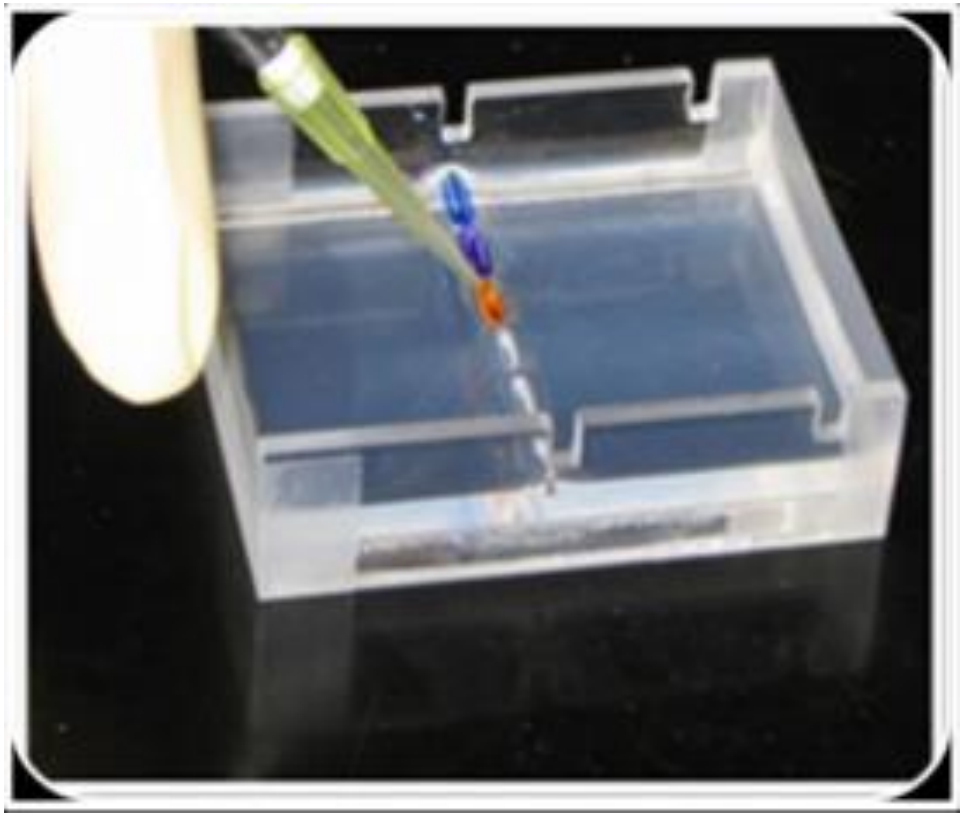
Agarose Gel Electrophoresis

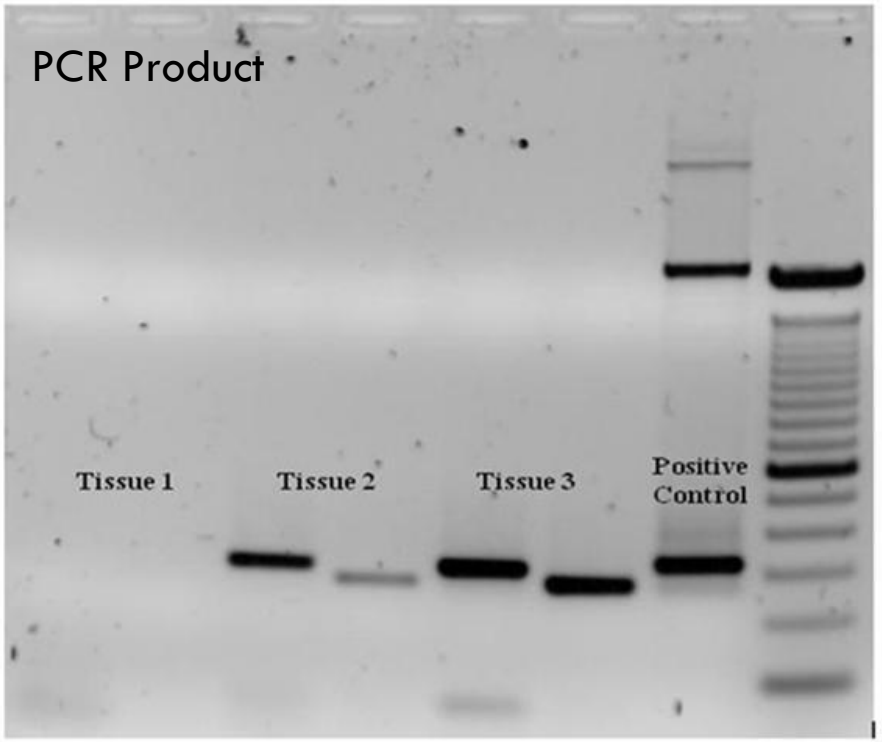
- It is a method used in biochemistry and molecular biology to **separate** DNA, or RNA molecules based upon charge, size and shape.

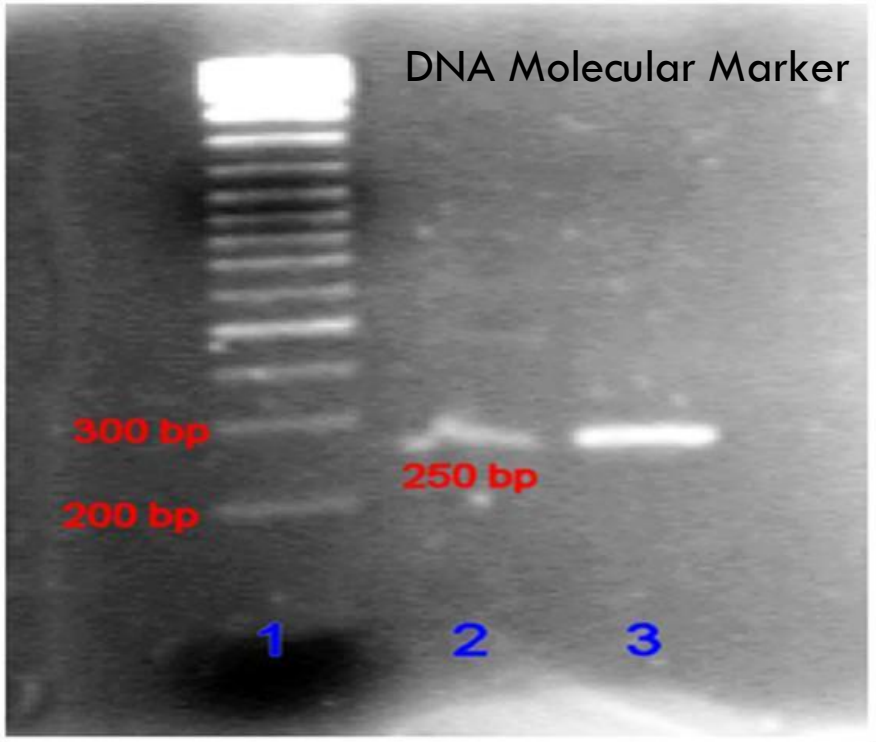
- Agarose is a polysaccharide derivative of agar.

Gel Tray/ Loading









- Amplified fragments can be visualized easily following staining with a chemical stain such as ethidium bromide.

- The DNA fragments are separated by charge and the relative sizes of fragments are determined by comparing to a standard DNA ladder.

» Factors, affect the mobility of molecules in gel

- Charge
- Size
- Shape
- Buffer conditions
- Gel concentration and
- Voltage

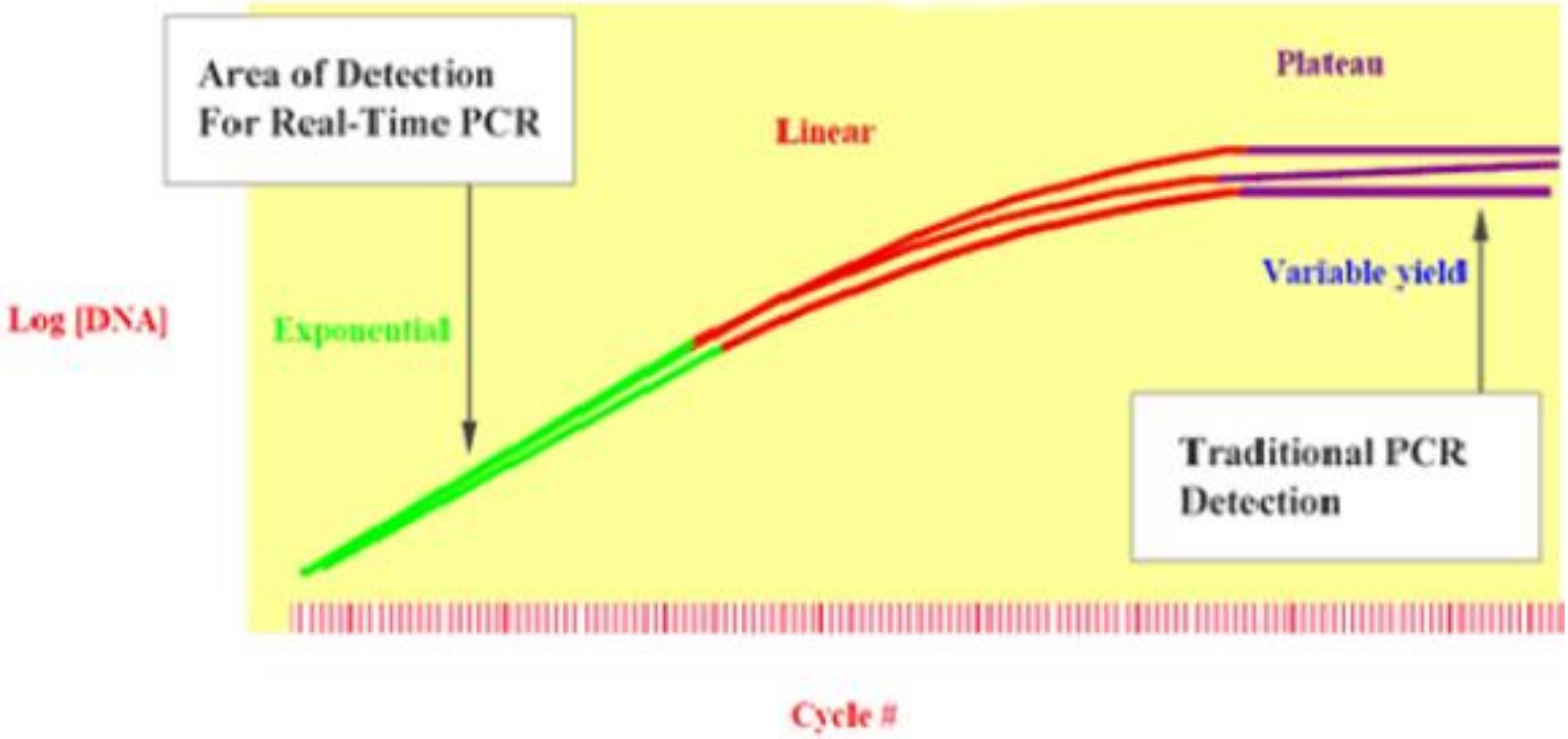
PCR: Three Phases

- Exponential: Exact doubling of product is accumulating at every cycle (assuming 100% reaction efficiency). The reaction is very specific and precise.

- Linear: The reaction components are being consumed; the reaction is **slowing**, and products are starting to degrade.

- Plateau: The reaction has **stopped**; no more products are being made and if left long enough; the PCR products will begin to degrade.

PCR Phases


Area of Detection
For Real-Time PCR
Plateau
Linear
Variable yield
Log [DNA]
Exponential
Traditional PCR
Detection
Cycle #

Polymerase Chain Reaction

Advantages of PCR

- Useful non- invasive procedure.
- Simplicity of the procedure.
- Sensitivity of the PCR

Disadvantages of PCR

- False positive results (cross contamination).
- False negative results

Variant PCR

- Reverse transcriptase-PCR.
- Nested-PCR.
- Hot-start PCR.
- Quantitative PCR.
- Multiplex-PCR.
- Mutagenesis by PCR.
- Allele specific PCR.
- …..

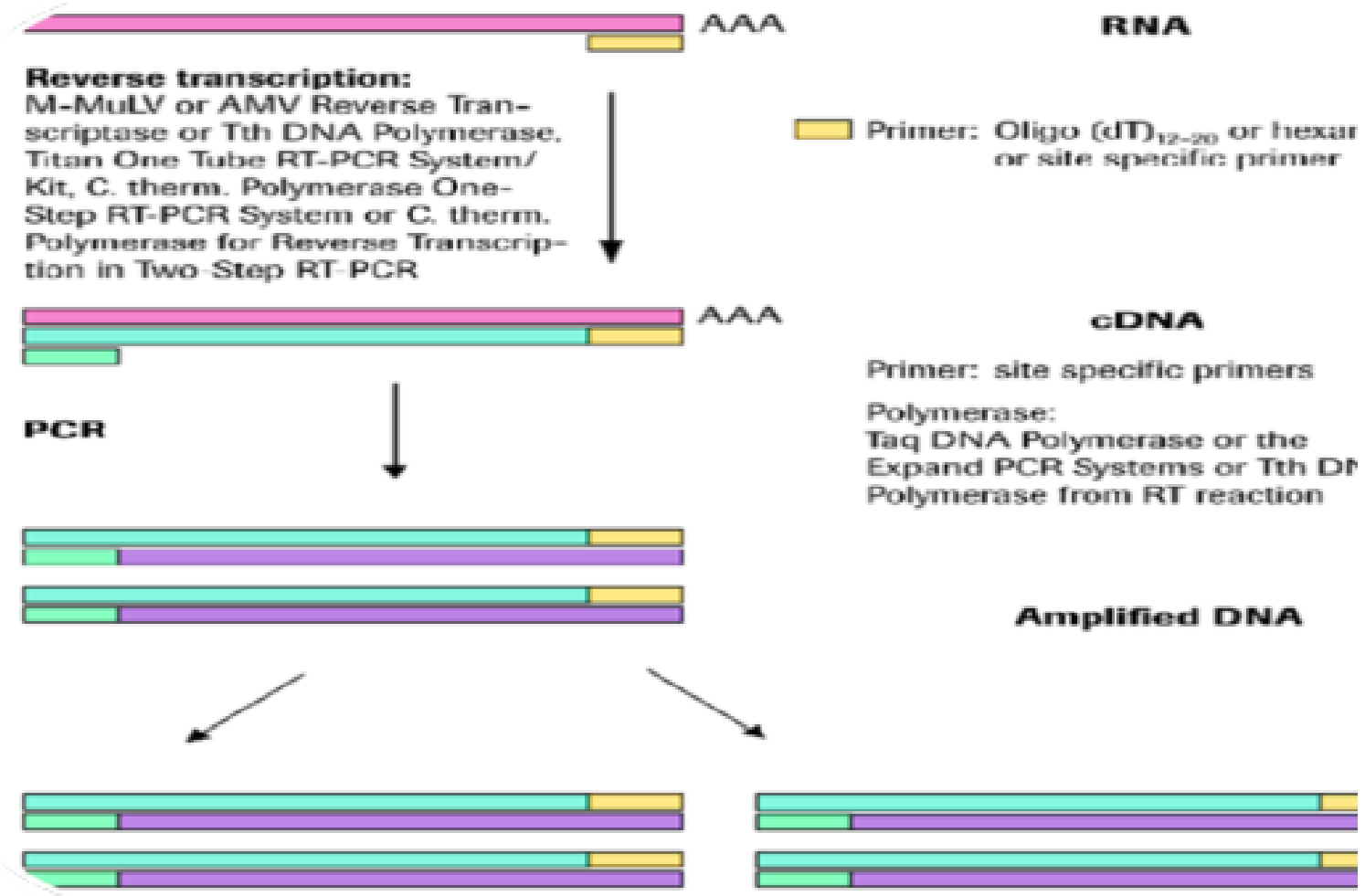
Reverse Transcriptase - PCR

RT-PCR, one of the most sensitive methods for the detection and analysis of rare mRNA transcripts or other RNA present in low abundance.

RNA cannot serve as a template for PCR.

RNA must be first transcribed into cDNA with reverse transcriptase from Moloney murine leukemia virus or Avian myeloblastosis virus, and the cDNA copy is then amplified.

RT- PCR

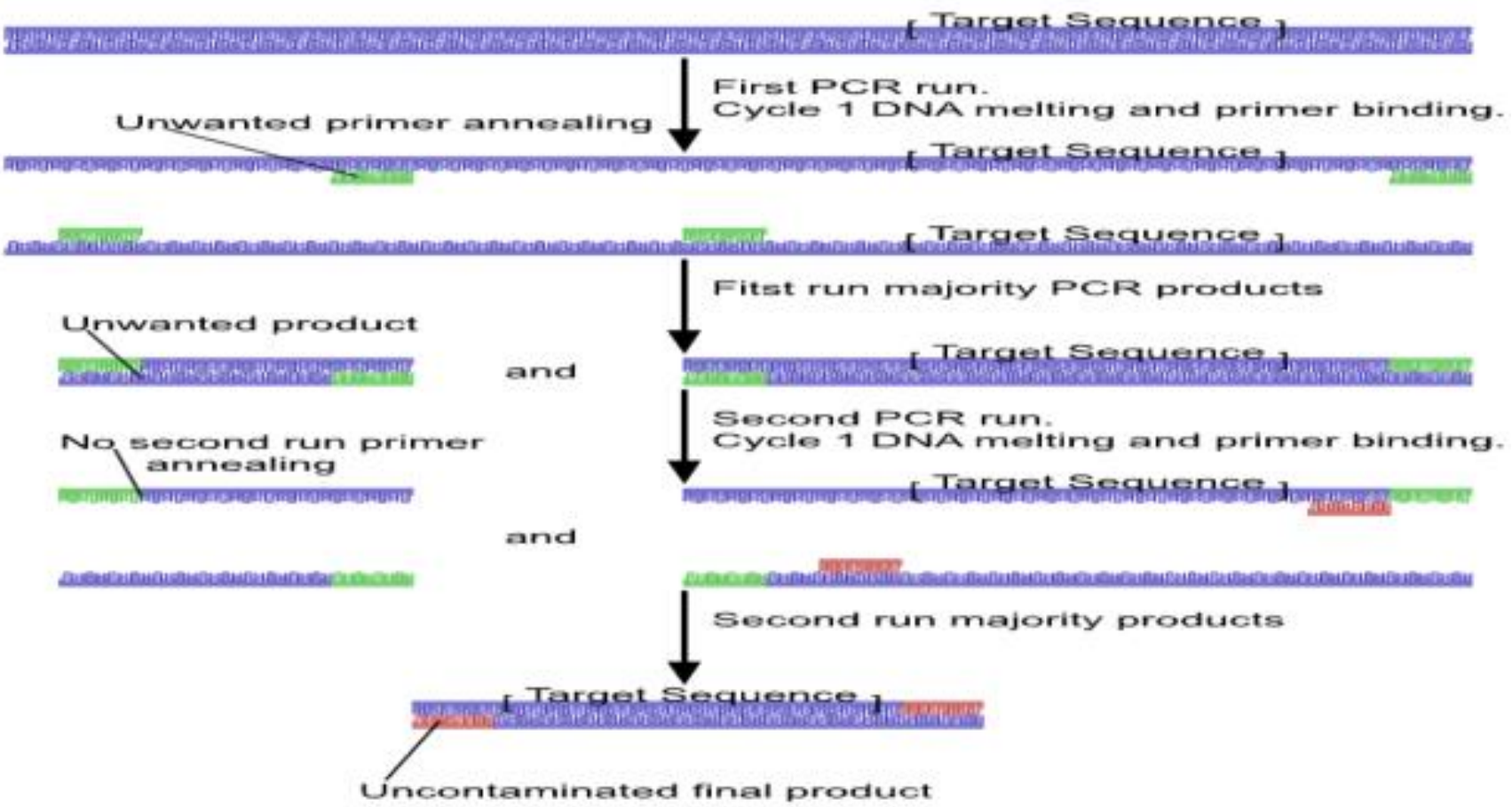

AAA
RNA
Reverse transcription:
M-MuLV or AMV Reverse Tran-
scriptase or Tth DNA Polymerase.
Titan One Tube RT-PCR System/
Kit, C. therm. Polymerase One-
Step RT-PCR System or C. therm.
Polymerase for Reverse Transcrip-
tion in Two Step RT-PCR
Primer: Oligo (dT)12-20 or hexar
or site specific primer
AAA
cDNA
PCR
Primer: site specific primers
Polymerase:
Taq DNA Polymerase or the
Expand PCR Systems or Tth DN
Polymerase from RT reaction
Amplified DNA

Nested PCR

- Nested PCR is a very specific PCR amplification.

- Nested PCR use two pairs (instead of one pair) of PCR primers are used to amplify a fragment.

Nested - PCR


Target Sequence
First PCR run.
Cycle 1 DNA melting and primer binding.
Unwanted primer annealing
Target Sequence
Target Sequence
Fitst run majority PCR products
Unwanted product
and
Target Sequence
Second PCR run.
Cycle 1 DNA melting and primer binding.
No second run primer annealing
Target Sequence
and
Second run majority products
Target Sequence
Uncontaminated final product

Hot - Start PCR

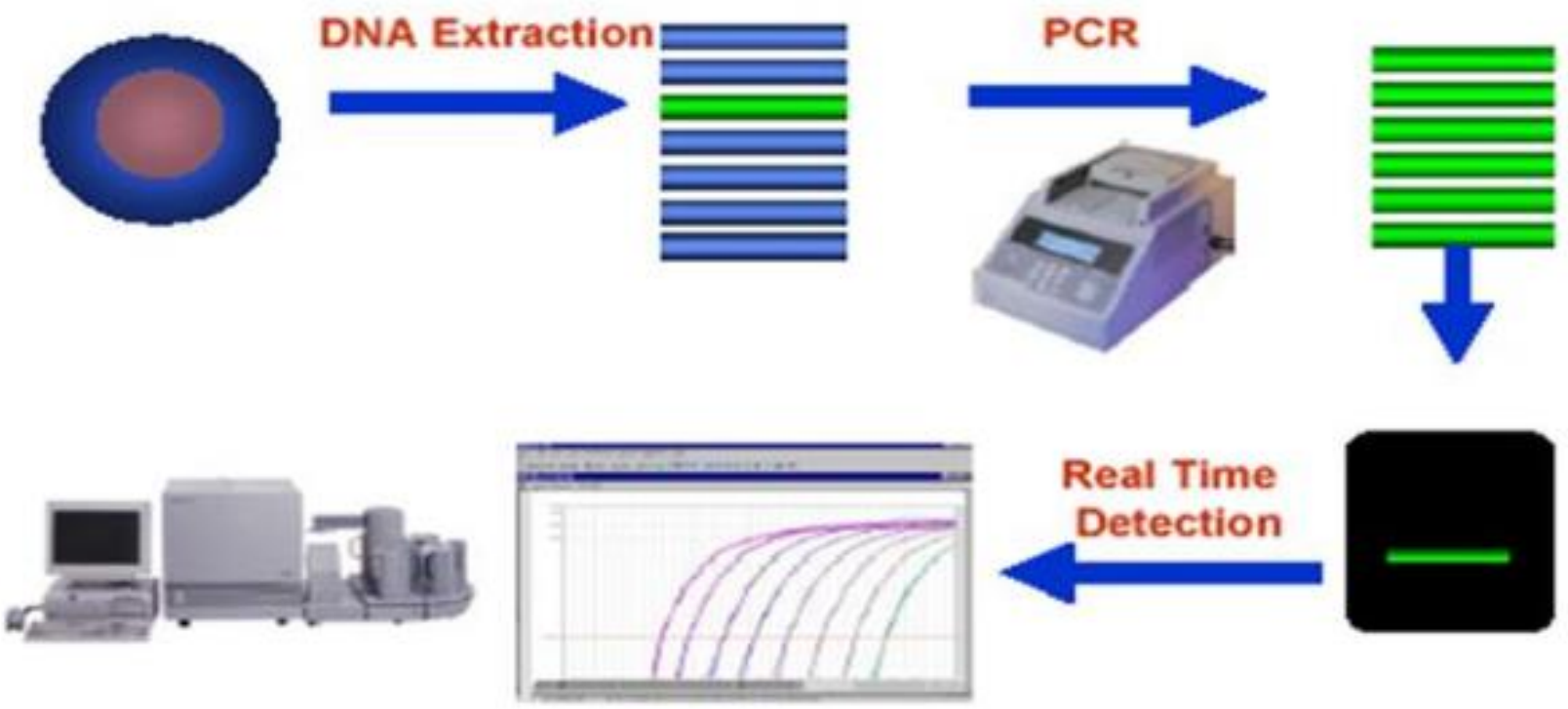
- Hot Start PCR significantly improves specificity, sensitivity and yield of PCR.

- The technique may be performed **manually** by **heating** the reaction components to the melting temperature (e.g., 95°C) before adding the polymerase. Specialized enzyme systems can be used.

Real Time PCR

- Traditional PCR has advanced from detection at the end-point of the reaction to detection while the reaction is occurring (Real-Time).
- Real-time PCR uses a fluorescent reporter signal to measure the amount of amplicon as it is generated . This kinetic PCR allows for **data collection after each cycle** of PCR instead of only at the end of the 20 to 40 cycles.

Real Time PCR



POLYMERASE CHAIN REACTION: USES

- Molecular biology,
- Microbiology,
- Genetics,
- Diagnostics clinical laboratories,
- Forensic science,
- Environmental science,
- ……

Real Time PCR : Uses