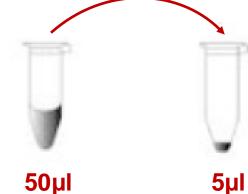


Dr. Nesrin Mwafi

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- Three routine tests can be performed to check for quantity (concentration/ amount / yield) and quality (purity and integrity) of the extracted product
- Take an aliquot of the sample (5µl out of 50µl)



**1. Measurement of concentration** 

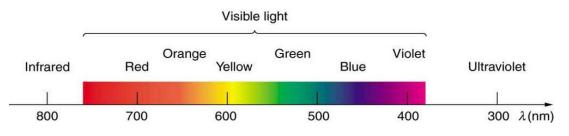




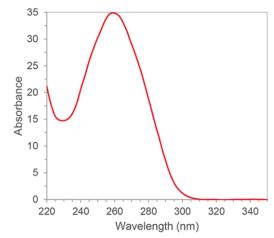


 UV-Vis spectrophotometer (ultraviolet-visible spectrophotometer) instrument is used to measure the concentration and the yield of the extracted nucleic acid (DNA or RNA)

## Electromagnetic Spectrum of Light



Nucleic acids absorb UV-light with maximum absorbance at wavelength of 260 nm  $(\lambda = 260 \text{ nm})$ 

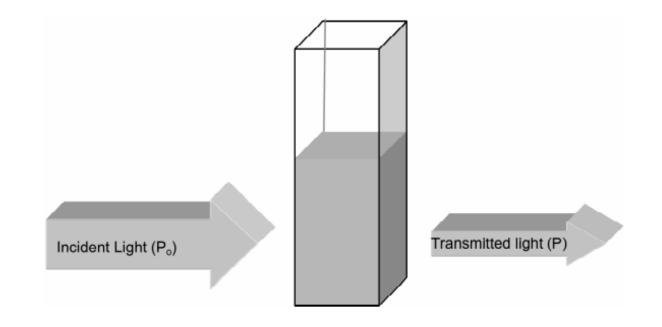


Typical RNA/DNA absorbance spectrum.

### **Assessment of Extracted Nucleic Acid** Monochromator Light Source Detector $I_t$ **|**\_ Entrance Dispersive Exit Slit element Sample Slit Wavelength selector



- Part of the incident light will be absorbed by the sample particles (depending on the concentration)
- The attenuation in the light that reaches the detector is measured in relation to the incident light and expressed as optical density (OD)

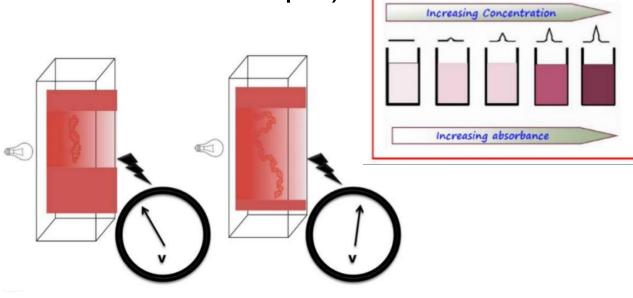




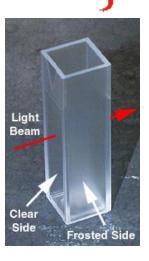
 The concentration is estimated per one optical density unit (10D) according to the type of nucleic acid (e.g. 1 OD unit = 50 µg/ml dsDNA)

Nucleic Acid	Concentration (µg/ml) or (ng/µl) per 1 OD Unit
ds DNA	50
ss DNA	33
ss RNA	40

- Cuvette is made of plastic (disposable) or quartz with two transparent and two opaque (foggy/frosted) sides
- Before sample measurement, a blank must be measured (the buffer or solvent used to dissolve the sample)









- If DNA conc. in a sample is 3000 ng/µl (or 3000 µg/ml or 3 µg/µl) then calculate the amount (yield) of DNA if total volume is 50 µl ?

**Answer:** Amount/ yield = concentration X total volume of the extracted sample

- = 3000 ng/µl X 50 µl
- = 150000 ng or 150 µg

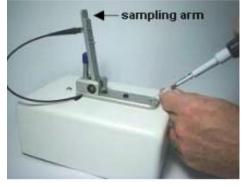
Note: 1g = 1000mg $1mg = 1000 \mu g$  $1 \mu g = 1000 ng$  1L = 1000 ml 1ml = 1000 μl



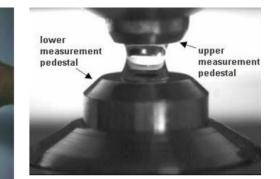


NanoDrop is automatically calculate the concentration in ng/ $\mu$ l

- NanoDrop spectrophotometer has similar principle to the standard cuvette spectrophotometer
- Load 1µl blank onto the lower pedestal, close the sampling arm and click on blank (wait 10-15 second)
- Wipe the pedestals and repeat the same steps using 1µl DNA or RNA sample and click on measure











## 2. Sample purity:

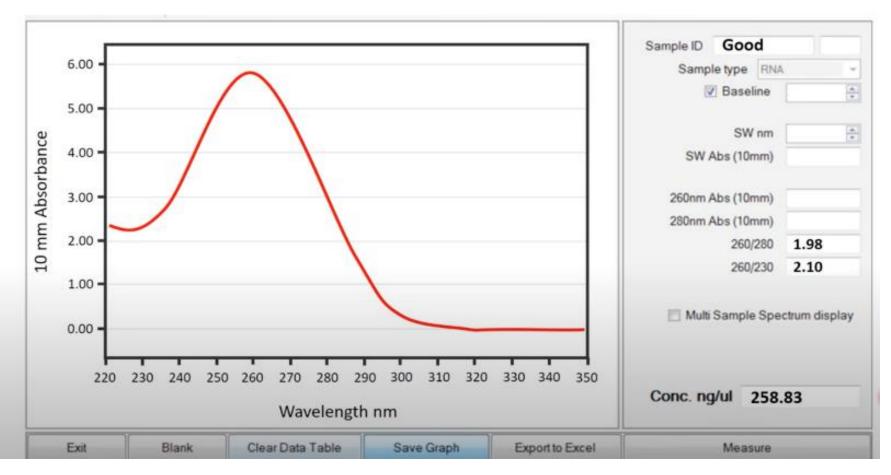
- The reading of absorbance at 260nm is divided by the reading at 280nm to estimate sample purity
- Aromatic amino acids of protein have maximum absorption at 280nm
- This ratio is most commonly used to determine the presence of protein in the isolated sample
- The acceptable range for this ratio:

Sample type	Ideal	Accepted range
DNA	1.8	1.7-1.9
RNA	2.0	1.9-2.1

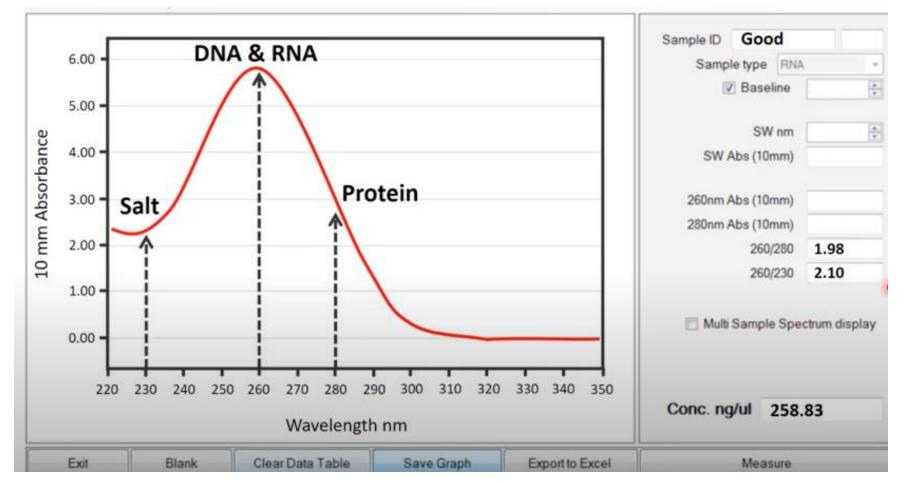


- Lower ratio indicates protein contaminants
- The ratio of A260/A230 is used as secondary measure of nucleic acid purity (chaotropic salts, TRIzol and peptide bonds of protein absorb at 230nm)
- The expected value is 2.0-2.2 (should be greater than A260/A280 ratio)

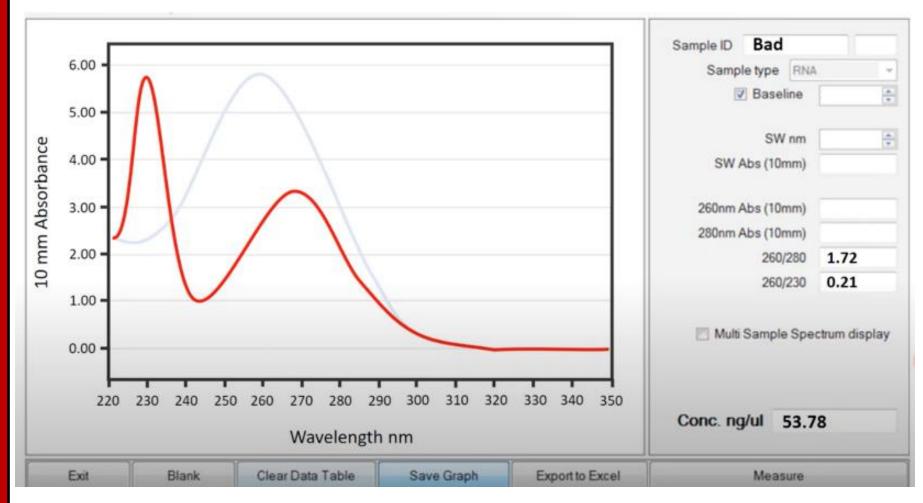




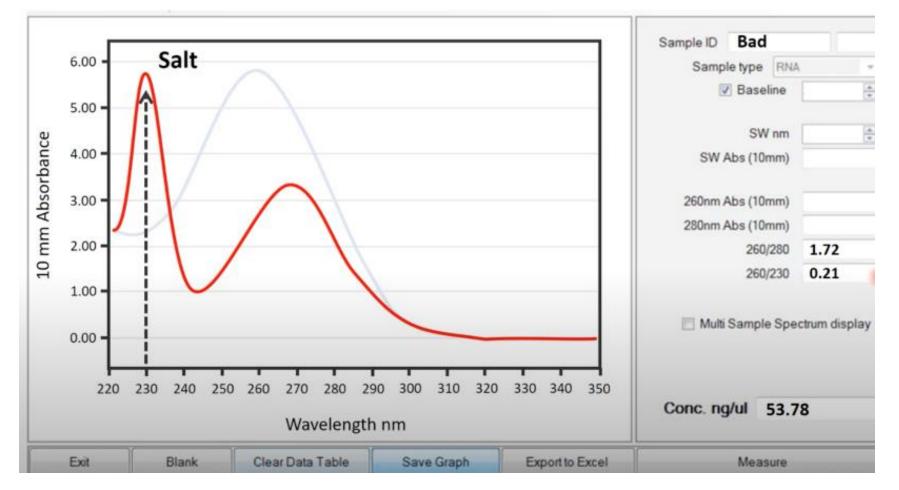




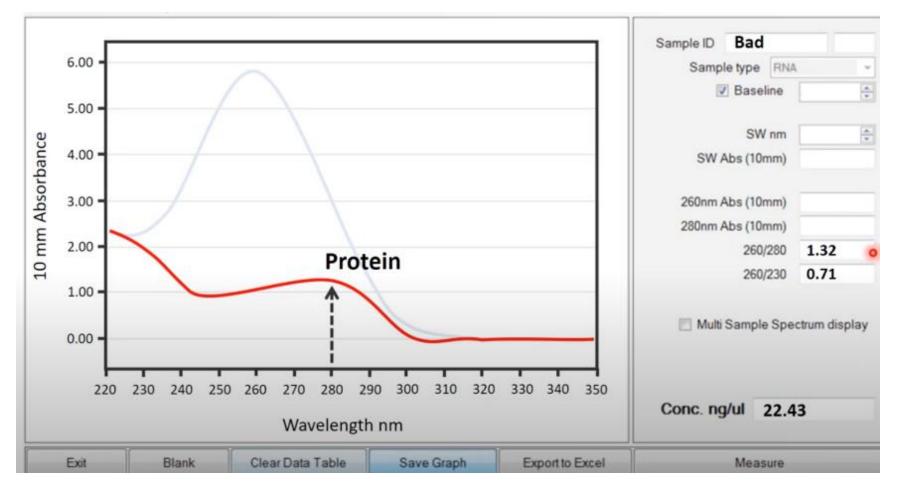








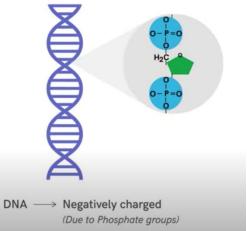






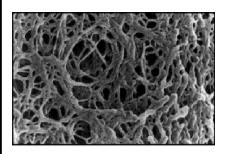
## 3. Gel electrophoresis:

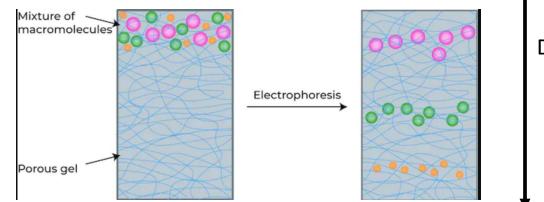
- Gel electrophoresis is a standard lab procedure for separation nucleic acids based on their sizes under the influence of electric field
- The concept: DNA and RNA are negatively charged molecules they move toward the positive electrode (usually red)





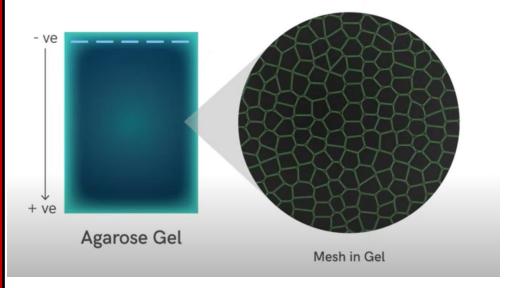
- Two types of gel can be used: polyacrylamide gel (suitable to separate small fragments up to 500bp) and agarose gel (suitable to separate larger fragments)
- Gel matrix acts as sieve or mesh (porous) and the smallest fragments migrate faster through the pores

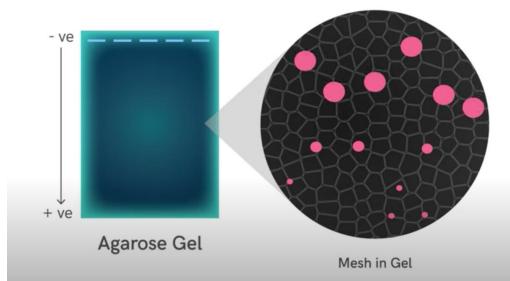




Direction of fragments migration





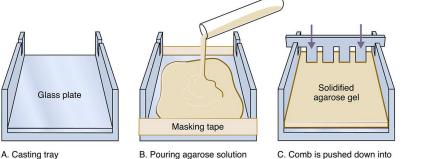




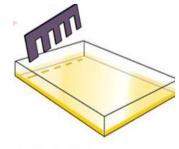
- For example: to prepare 1% agarose gel (1g/100ml) dissolve 1g of agarose powder in100ml of buffer
- add few drops of nucleic acid fluorescent dye to the dissolved gel solution then pour it into casting tray (don't forget to add the comb to make the

ael to form wells

wells)

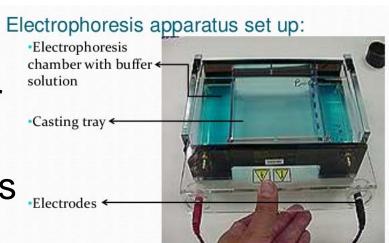


onto glass plate



3. Remove comb. Shallow wells are left in gel.

- The nucleic acid fluorescent dye is used to visualize the nucleic acid under UV light (acts by intercalation): Ethidium Bromide (mutagen). GelRed (expensive but safe)
- The gel tray is placed in an electrophoresis chamber and filled with running buffer until it covers the gel piece. Buffer is used to provide ions that carry the current and to maintain pH

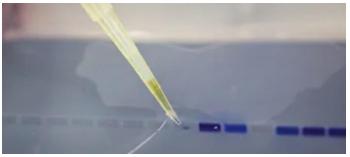


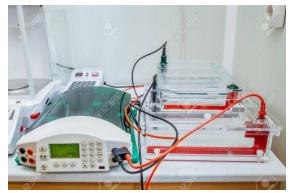
ethidium bromide (marked red)

intercalates between base pairs

- Load the DNA or RNA sample into wells after mixing with loading dye (blue dye to increase viscosity of sample and prevents it from floating out of the wells and to track the migrated fragments)
- The electrodes are attached to a power supply and an electrical current is applied



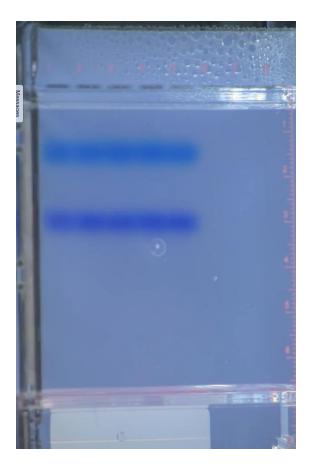












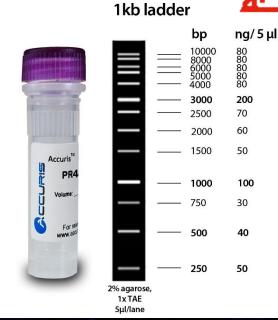
#### After 5 min

After 20 min

 Load DNA or RNA ladder (fragments with known sizes) into the first well (acts a ruler to compare and identify sizes of different bands)

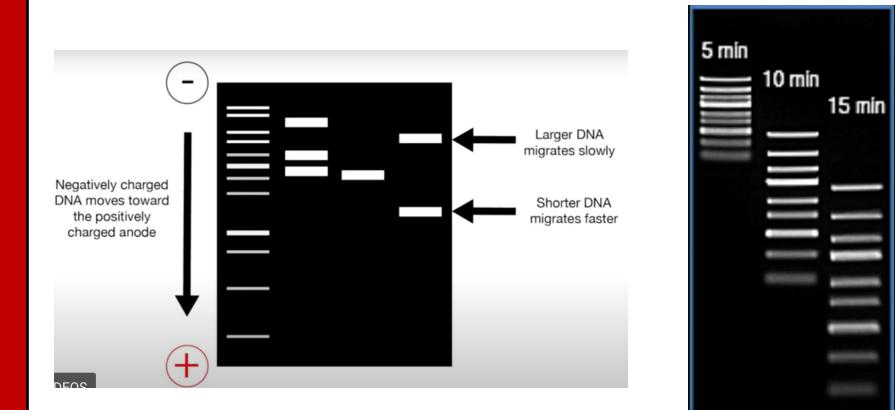


Gel documentation system "Gel Doc System"

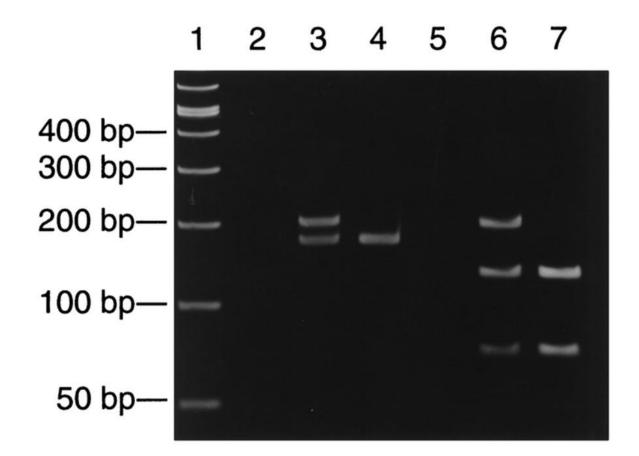






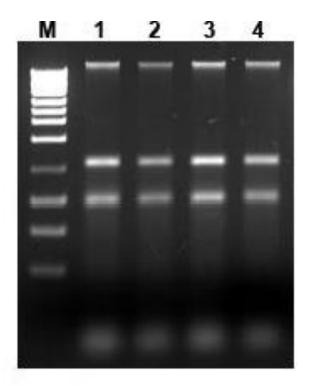




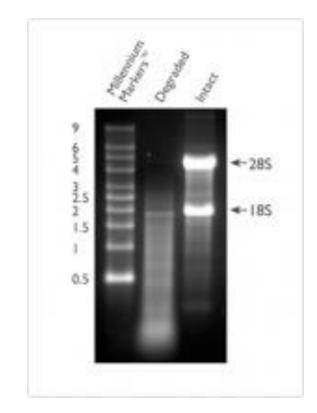


## **Assessment of Extracted RNA**

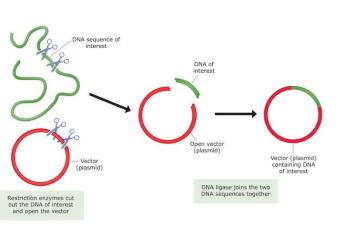




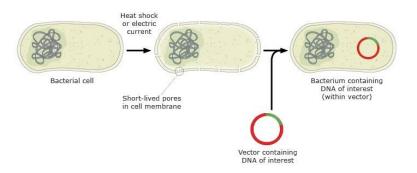
Total RNA



## **Assessment of Extracted plasmid**



1. Ligation of gene of interest with plasmid



#### 2. Bacterial transformation for amplification

Non-linearized plasmid



Plasmid linearization by restriction enzyme with unique cut site at gene of interest

# Assessment of Extracted Plasmid

